

Tetrahedron: Asymmetry 13 (2002) 2471-2483

Stereoselective synthesis of epoxyalkyl glycoside precursors of glycosyl glycerol analogues from alkenyl glycosides of N-acetyl-D-glucosamine derivatives[†]

José M. Vega-Pérez,* José I. Candela, Eugenia Blanco and Fernando Iglesias-Guerra*

Departamento de Química Orgánica y Farmacéutica, Facultad de Farmacia, Universidad de Sevilla, E-41071 Sevilla, Spain

Received 23 September 2002; accepted 10 October 2002

Abstract—The synthesis of epoxyalkyl glycoside derivatives of N-acetyl-D-glucosamine is described. Epoxidation of the corresponding alkenyl glycosides with *m*-CPBA took place with different stereoselectivity depending on the nature of the unsaturated system and the protecting groups on the sugar moiety. The configuration of the newly formed stereogenic centres has been confirmed unequivocally by chemical correlation. © 2002 Elsevier Science Ltd. All rights reserved.

1. Introduction

Oxiranes are important intermediates in organic synthesis, because their electrophilic or nucleophilic opening leads to 1,2-difunctionalized systems or to the formation of new carbon-carbon bonds.²⁻⁴ This chemical reactivity is responsible for the biological activity exhibited by compounds bearing an oxirane moiety. Thus, they can present cytotoxic activity, and hence have been studied for use as anti-cancer agents.⁵ It is noteworthy that in a strategy to obtain new highly selective drugs against carcinoma cells in rapid growth, which have a great demand for primary metabolites such as carbohydrates, the latter are used as antitumour agent carrier.^{6–8} This line of research is one of the components of our work, and previously we have described the synthesis of compounds with potential antitumour activity by linkage of the alkylating moiety-chlorambucil or cyclophosphamide-widely used clinically in the treatment of different types of cancer^{9,10}—to different positions of various aminosugars.^{1,11–14} We want to broaden this approach and use oxiranes as alkylating moiety. At the same time, the epoxyalkyl glycosides of some sugars have been studied as potential inhibitors of several endoglycosidases. Thus, epoxypropyl glycosides of N-acetyl-D-glucosamine, di-N-acetylchitobiose and tri-*N*-acetylchitotriose have been prepared as inhibitors

of lysozyme,15 while epoxyalkyl cellobiosides have been synthesized as inhibitors of cellulases.^{16,17} It has also been reported that the inhibition of several endoglucanases by oligosaccharidic epoxyalkyl glycosides shows a remarkable specificity not only with respect to the sugar moiety but also with respect to the chain length and the configuration of the aglycone.^{18,19} Furthermore, it is noteworthy that certain glycosyl glycerols, an important class of compounds widely distributed in plants, animals and bacteria,²⁰ have shown tumour inhibitory activity;^{21–25} and they can be obtained from epoxyalkyl glycosides and subsequent ring opening reaction.¹⁶ Thus, we became very interested in investigating a good method to obtain the epoxyalkyl glycosides of aminosugars with high stereoselectivity.

We believe that a straightforward means to achieve this might be the oxidation of alkenyl glycosides, for the following (and other) reasons.

(a) Alkenyl glycosides are readily obtained, and the oxidation of alkenes to oxiranes is very well known and easily performed;

(b) published precedent works on the oxidation of alkenyl glycosides are scarce, and refer mostly to allyl glycosides of sugars;^{26–28} those referring to the epoxidation of alkenyl glycosides derived from *N*-acetyl-D-glucosamine are even fewer in number;²⁹ and

(c) we have used 2-aminosugars as chiral templates in the stereoselective synthesis of numerous compounds (2-aminoglycals,³⁰ 2-nitrosugars,¹⁴ chiral oxazolidi-

^{*} Corresponding authors. Tel./fax: +34 95 4556737; e-mail: vega@fafar.us.es, vega@us.es

[†] Potential anticancer drugs, Part 6. For Part 5, see: Ref. 1.

^{0957-4166/02/\$ -} see front matter @ 2002 Elsevier Science Ltd. All rights reserved. PII: S0957-4166(02)00649-3

nes,³¹ chiral epoxyamides,³² compounds with potential anti-cancer activity^{11–13}), and we anticipated that the glucosamine moiety could serve as a chiral inductor in the oxidation reaction and as carrier agent in the derivatives with potential biological activity.

We have already published preliminary assays for the preparation of epoxyalkyl glycosides by oxidation of alkenyl glycosides of D-glucosamine,¹ and observed that the reaction takes place with high stereoselectivity. We have also explored the preparation of glycosyl amino-glycerols by further ring opening reaction with nitrogen nucleophiles.³³

Herein, we report on a study of the oxidation of a wide range of alkenyl glycosides derived from D-glucosamine with *m*-chloroperoxybenzoic acid (*m*-CPBA), aiming to analyze the influence that the different groups on the double bond and the protecting groups of the sugar hydroxyls have on the stereoselectivity of the reaction. We also present unequivocal chemical proof of the configuration of the major diastereoisomer obtained in the oxidation reaction. The new compounds obtained can be converted into new glycosyl glycerols representing a pool of compounds, of differing—but related structure, with a potentially broad spectrum of biological activity.

2. Results and discussion

The oxidation of the alkenyl glucopyranosides 1-18 was studied (Schemes 1 and 2) in an attempt to determine the effect of their different structural features on the stereofacial differentiation of the diastereotopic faces of the double bond in the epoxidation reaction. They can be divided into two groups, according to their higher or lower conformational flexibility: the peracetylated derivatives 1, 2, 4, 5, and those possessing a benzylidene group between positions 4 and 6 of the tetrahydropyran ring, resulting in a more rigid structural model 7-18. This was important to study because the only work published on oxidations of the alkenyl glycosides of D-glucosamine deals solely with the oxidation of $1,^{29}$ and we ourselves have observed greater stereoselectivity in the oxidation of the more rigid compounds.³³ Because of the possible influence of the group at position 3 of the sugar on the stereochemical course of the reaction, we have further divided the benzylidene derivatives into two groups: compounds with the C-3 hydroxyl group free (7-12), and compounds with the C-3 hydroxyl group blocked with protecting groups, in analogy to the peracetylated derivatives (13-18). Finally, within each group, the glycosides are differentiated by the number and type of substituents on the double bond.



Scheme 1. Reagents: (i) AcCl; (ii) Unsaturated alcohol/Hg(CN)₂; (iii) NaMeO/MeOH; (iv) PhCHO/ZnCl₂.



Scheme 2. *Reagents*: (i) Ac₂O/Py; (ii) MeI/KOH/18-crown-6/THF or BnBr/CH₂Cl₂/50% NaOH/H₂O/TBABr; (iii) TBSCl/imida-zole/DMF; (iv) CH₂Br₂/CH₂Cl₂/50% NaOH/H₂O/TBABr/reflux.

The alkenyl glucopyranoses 1–12 were obtained from *N*-acetyl-D-glucosamine in excellent yields using the procedure described previously for alkyl glucopyranosides^{11,12,34} (Scheme 1). The compounds 1, 6 and 7 have been described previously in the literature.^{15,35,36} The reaction of 2-acetamide-3,4,6-tri-O-acetyl-2-deoxy- α -D-glucopyranosyl chloride³⁷ with the corresponding unsaturated alcohol gave compounds 1-6, which were transformed into the compounds 7-12 by deacetylation, and subsequent reaction with benzaldehyde. The NMR spectra showed signals corresponding to the unsaturated moiety incorporated into the sugar molecule in the glycosidation reaction. The ¹H NMR spectra showed signals at 6.60-4.80 ppm corresponding to the double bond, the characteristic signals corresponding to the sugar moiety, a doublet for H-1 at 4.70–4.50 ppm for the compounds 1–12, two triplets for H-3 and H-4 at 5.20 and 4.90 ppm, respectively, for 1-6, and a singlet at 5.50 ppm corresponding to the benzylidene group in compounds 7–12.

The syntheses of compounds 13-18 were carried out from 7 or 8 as indicated in Scheme 2. The ¹H NMR spectra of these compounds showed the signals corresponding to the protecting group present: a singlet at 2.07 ppm for 13, a singlet at 3.35 ppm for 14, two signals at 4.70-4.50 ppm for 15, three singlets at 0.79, -0.03 and -0.10 for 16 (similar for 17), and two signals at 5.48 and 4.80-4.70 for 18.

The reaction of 1, 2, 4, 5, 7–18 with *m*-CPBA in dichloromethane at different temperatures gave the corresponding oxiranes 19–34. In all cases, the major isomer was isolated and purified by recrystallization from ethanol or by flash chromatography on silica gel (Scheme 3, Table 1). The ¹H NMR spectra for these compounds showed the characteristic signals of the oxirane system at 3.85 and 3.20 ppm for 21 and 26, and at 3.10–2.50 for the other compounds (Table 2). The ¹³C NMR spectra showed signals between 62 and 44 ppm for every one the compounds 19–34.

The diastereomeric excess (d.e.), determined using ¹H NMR, is shown in Table 1, from which certain inferences can be drawn. For their analysis, the starting alkenyl glycosides can be grouped into the three groups defined above. The first comprises the peracetylated derivatives (compounds 1, 2, 4, 5); the second is the benzylidene derivatives with the 3-hydroxyl group free (compounds 7-12); and the third is the benzylidene derivatives with the 3-hydroxyl group blocked (compounds 13–18). As expected, the d.e. of the oxidation reaction is higher when the reaction temperature is lowered. The d.e. obtained for the compounds of the first group (entries 1–8) are lower than those for the corresponding compounds of the second group with the same aglycone (entries 9-20) measured at the same temperature, probably because of the greater rigidity of the latter compounds. So, at 10°C, compounds 23, 24, 26 and 27 are obtained with d.e. of 93, 82, 70 and 49%, respectively, while compounds 19, 20, 21 and 22 are obtained with d.e. of 72, 74, 60 and 34%, respectively. The d.e. of the alkenyl glycosides of the third group (entries 21-26) are again lower than those for the corresponding compounds of the second group with the same aglycone (entries 10 and 12) measured at the same temperature. This effect is very marked for the oxidation of 18, a compound in which, besides not having the hydroxyl group at position 3 of the sugar, also does not have the N-H bond by formation of an oxazolidine bridge between these two positions. This supports the idea of the formation of a hydrogen bond between the NH group and the peroxyacid, preferentially stabilizing the transition state related to electrophilic addition to the Re-face (see below) of the double bond,³³ and further suggests additional stabilization of this transition state, probably by formation of a hydrogen bridge between the hydroxyl group at position 3 and the acetamido group. At the same time, intra-group comparison of the different alkenyl glycosides by the degree of substitution of the double-bond carbons shows that diastereoselectivity generally decreases slightly when a



Scheme 3. *Reagents*: (i) *m*-CPBA/CH₂Cl₂.

Entry	Starting compound	Temperature (°C)	Reaction time ^a (days)	Reaction product	Yield ^b (%)	D.e. ^c (%)
1	1	10	5	19 ^d	75	72
2	1	-15	55	19	42	80
3	2	10	4	20	68	78
4	2	-15	60	20	57	95
5	4	10	6	21	69	60
6	4	-15	80	21	67	88
7	5	10	6	22	71	35
8	5	-15	90	22	52	43
9	7	10	2	23	74	93
10	7	-15	90	23	48	100
11	8	10	6	24	69	86
12	8	-15	33	24	49	97
13	9	10	1	25	81	66
14	9	-15	30	25	66	82
15	10	10	2	26	72	75
16	10	-15	24	26	65	87
17	11	10	6	27	73	49
18	11	-15	70	27	58	61
19	12	10	2	28	70	27
20	12	-15	35	28	59	36
21	13	-15	20	29	82	70
22	14	-15	60	30	47	74
23	15	-15	14	31	85	68
24	16	-15	30	32	83	64
25	17	-15	14	33	76	74
26	18	-15	72	34	62	23

Table 1. Epoxidation of alkenyl glucosides 1, 2, 4, 5, 7–18 with m-CPBA

^a Reaction time approximated (TLC showed that all starting compound had been consumed).

^b Yields refer to compounds obtained in each reaction after isolation and purification.

^c Determined by relative integration in the ¹H NMR spectra of reaction mixtures.

^d Literature: temperature 20°C; yield 71%; d.e. 68% (Ref. 29).

Table 2. NMR data (δ , ppm) in CDCl₃ for the oxirane protons, and anomeric proton and carbon in compounds 19–34



Entry	Compound	H-2′	H _A -3'; H _B -3'	H-1 minor/major	C-1 major/minor
1	19 ª	3.12	2.75; 2.67	4.75/4.63	
2	19	3.12	2.76; 2.68/2.53 ^b	4.74/4.62	101.3/100.7
3	20	_	2.82; 2.55	4.67/4.54	101.6/101.2
4	21	3.17	3.85; -	4.81/4.65	101.6/100.8
5	22	2.92	-; -	4.79/4.70	101.8/100.2
6	23 ^{c,d}	3.05	2.70; 2.56	4.54/4.51	101.8/101.6
7	24	_	2.95; 2.66	4.67/4.44	102.9/102.8
8	25	3.10	2.89; -	4.68/4.49	101.7/101.6
9	26 ^d	3.24	3.85; -	4.59/4.55	102.0/101.9
10	27	2.99	-; -	4.75/4.67	101.7/101.3
11	28	3.06	2.81; 2.56	4.62/4.53	101.7/101.4
12	29	3.06	2.76; 2.64/2.51 ^b	4.55/4.43	102.5/102.1
13	30	3.11	2.77; 2.65/2.55 ^b	5.05/4.95	101.2/101.1
14	31	3.10	2.76; 2.64/2.52 ^b	4.96/4.9-4.8	101.0/100.7
15	32	3.11	2.76; 2.66/2.52 ^b	4.98/4.90	100.8/100.5
16	33	_	2.81; 2.57	4.92/4.80	101.2/100.8
17	34 ^{d,e}	3.15	2.73; 2.50	4.96/4.95	103.5/103.3

^a Reference 29.

 $^{\rm b}\,\delta\,$ $\rm H_{B}$ for the major/minor isomers.

^c Reference 33.

^d NMR spectrum recorded in DMSO-*d*₆.

^{e 1}H NMR spectrum recorded at 80°C.



Scheme 4. Reagents: (i) NaN₃/LiClO₄/CH₃CN; (ii) NaMeO/MeOH; (iii) PhCHO/ZnCl₂; (iv) Ac₂O/Py.

single substituent is introduced at the unsaturated carbons (see, for example, the entries for compounds 24-26 with respect to 23) and decreases notably when the allylic system is doubly substituted (see the entry for 22 with respect to 19, and 27 with respect to 23). Finally, when the aglycone is a homoallylic system (compound 12, entries 19 and 20), the oxidation product 28 is obtained with very low stereoselectivity.

In conclusion, the structural features favouring high stereoselectivity in the oxidation of alkenyl glycosides of N-acetyl-D-glucosamine derivatives by m-CPBA, are, probably in this order: the fact that the alkenyl glycoside is an allyl glycoside; the existence of the N–H bond in the acetamido group; the presence of an acetal group between positions 4 and 6 of the pyranose ring, making the system rigid; and the presence of a free hydroxyl group at position 3. The existence of all these features in compound 7 explains why, at low temperature, its oxidation product—compound 23—can be obtained with total diastereoselectivity.

Regarding the stereochemistry of the new stereogenic centre created in the formation of the oxirane, Peter et al.²⁹ demonstrated unequivocally that, in compound **19** (the major diastereoisomer), the configuration is R at position 2' (see Table 2). They also observe that the anomeric proton of this major compound **19** (C-2' R) resonates at higher field (or possesses a lower δ) than the minor diastereoisomer, (C-2' S), and on this basis, assign the R-configuration for the other oxiranes (different to those presented here) described in their work.

In all of the cases examined (as shown in Table 2) the major stereoisomer also presents a lower chemical shift in ¹H NMR for the value of the anomeric proton (as in **19**) and a higher chemical displacement value for the anomeric carbon in ¹³C NMR. Thus, it can be tentatively stated that the *Re*-face of the double bond is the more reactive to the electrophilic agent.

To rule out the possibility that the presence of the benzylidene ring condensed with the tetrahydropyran ring in compounds 7–18 might invert the reactivity of the diastereotopic faces of the alkene and the chemical displacement of the anomeric protons of the oxiranes

formed (compounds 23–34), we went on to the chemical assignment of the stereochemistry of one of the benzylidene oxiranes (23) of defined stereochemistry (by correlation here with 19).²⁹ Thus, 36 was synthesized independently from 19 and 23³³ (Scheme 4). The chemical displacement values shown in its ¹H and ¹³C NMR spectra were identical to the signals for the major diastereoisomer. Both compounds 36 were transformed independently into 37; again, the NMR spectra were identical. The ¹H and ¹³C NMR spectra of a mixture of compound 37 obtained by the two routes present a single set of signals. The foregoing proves the *R*configuration at position 2' of 23, and by extension, recognizes the *Re*-face as the more reactive in all cases.

3. Experimental

3.1. General

Evaporations were conducted under reduced pressure. Preparative chromatography was performed on Silica Gel 60 (E. Merck). Kieselgel 60 F_{254} (E. Merck) was used for TLC. Melting points are uncorrected. Optical rotations were obtained on a Bellingham+Stanley Ltd P-20 polarimeter at 25°C. Mass spectra were recorded on a Micromass AUTOSPECQ mass spectrometer, CI at 150 eV, and HR mass measurements with resolutions of 10000. FAB mass spectra were recorded on a Kratos MS-80-RFA using a thioglycerol matrix. NMR spectra were recorded at 25°C on a Bruker AC-200 spectrometer at 200 MHz for ¹H and 50 MHz for ¹³C, and on a Bruker AMX-500 spectrometer at 500 MHz for ¹H and 125 MHz for ¹³C. The chemical shifts are reported in ppm on the δ scale relative to TMS. COSY, DEPT, and CHCORR experiments were performed to assign the signals in the NMR spectra.

3.2. Alkenyl 2-acetamido-3,4,6-tri-*O*-acetyl-2-deoxy-β-D-glucopyranosides, 1–6

A solution of 2-acetamido-3,4,6-tri-O-acetyl-2-deoxy- α -D-glucopyranosyl chloride³⁷ (18.3 g, 50 mmol) in dry nitromethane (40 mL) was added dropwise, with exclusion of moisture, to a stirred mixture of the corresponding unsaturated alcohol (25 mL) in anhydrous toluene (40 mL) containing 4 Å molecular sieves (30 g) and mercury(II) cyanide (12.5 g). The mixture was stirred at room temperature for 1–2 days. When TLC showed that all of the chloride had reacted, the mixture was diluted with ethyl acetate and filtered through a pad (12 cm diameter×1 cm height) of alumina. The organic layer was washed with an aqueous saturated solution of sodium bicarbonate and water, then dried (Na₂SO₄) and concentrated. Suspension of the residue with stirring in hexane for 1 h gave a solid that was filtered off and washed with hexane. Recrystallization from ethanol gave the pure peracetate.

3.2.1. 2-Methyl-2-propenyl 2-acetamido-3,4,6-tri-O-acetyl-2-deoxy-β-D-glucopyranoside, 2. Yield 17.0 g (85%); mp 165–166°C; $[\alpha]_D$ +36.9 (*c* 0.6, CH₂Cl₂); MS (CI): *m*/*z* 402 (65%) [M+H]⁺. ¹H NMR (200 MHz, CDCl₃): δ 5.84 (d, 1H, $J_{2,\text{NH}}$ 8.9 Hz, NH), 5.24 (t, 1H, $J_{2,3} = J_{3,4}$ 9.4 Hz, H-3), 5.02 (t, 1H, $J_{3,4} = J_{4,5}$ 9.5 Hz, H-4), 4.90, 4.85 (2bs, 2H, C=CH₂), 4.60 (d, 1H, J_{1,2} 8.3 Hz, H-1), 4.3-3.8 (m, 5H, H-2, H-6, H-6', OCH₂R), 3.65 (m, 1H, H-5), 2.03, 1.98, 1.97, 1.89 (4s, 4CH₃CO), 1.66 (s, 3H, CH₃). ¹³C NMR (50 MHz, CDCl₃): δ 170.9, 170.7, 170.2, 169.3 (4C=O), 140.9 (C=CH₂), 113.2 (C=CH₂), 99.5 (C-1), 72.8 (OCH₂R), 72.4 (C-4), 71.6 (C-3), 68.7 (C-5), 62.1 (C-6), 54.5 (C-2), 23.2 (CH₃CON), 20.7, 20.6, 20.5 (3CH₃COO), 19.2 (CH₃). HRMS (EI): [M]^{+•}, found 401.167637. C₁₈H₂₇NO₉ requires 401.168582. Anal. calcd for C₁₈H₂₇NO₉: C, 53.86; H, 6.78; N, 3.49. Found: C, 53.81; H, 6.67; N, 3.53%.

3.2.2. *trans*-2-Butenyl 2-acetamido-3,4,6-tri-*O*-acetyl-2deoxy-β-D-glucopyranoside, **3.** Yield 13.6 g (68%); mp 163–164°C; $[\alpha]_D = 28.3$ (*c* 0.8, CH₂Cl₂); MS (CI): *m/z* 402 (40%) [M+H]⁺. ¹H NMR (200 MHz, CDCl₃): δ 5.8–5.6 (m, 3H, NH, CH=CH), 5.28 (t, 1H, $J_{2,3} = J_{3,4}$ 9.6 Hz, H-3), 5.02 (t, 1H, $J_{3,4} = J_{4,5}$ 9.4 Hz, H-4), 4.69 (d, 1H, $J_{1,2}$ 8.3 Hz, H-1), 4.3–3.9 (m, 4H, H-6, H-6', OCH₂R), 3.80 (m, 1H, H-2), 3.66 (m, 1H, H-5), 2.05, 1.99, 1.98, 1.91 (4s, 4CH₃CO), 1.67 (d, 3H, *J* 6.5 Hz, CH₃). ¹³C NMR (50 MHz, CDCl₃): δ 170.8, 170.7, 170.3, 169.4 (4C=O), 130.7, 126.2 (CH=CH), 99.2 (C-1), 72.3 (C-4), 71.6 (C-3), 69.8 (OCH₂R), 68.6 (C-5), 62.1 (C-6), 54.7 (C-2), 23.3 (CH₃CON), 20.7, 20.6, 20.5 (3CH₃COO), 17.8 (CH₃). HRMS (CI): [M+H]⁺, found 402.176338. C₁₈H₂₈NO₉ requires 402.176407. Anal. calcd for C₁₈H₂₇NO₉: C, 53.86; H, 6.78; N, 3.49. Found: C, 53.53; H, 6.79; N, 3.48%.

3.2.3. trans-3-Phenyl-2-propenyl 2-acetamido-3,4,6-tri-Oacetyl-2-deoxy-β-D-glucopyranoside, 4. Yield 18.7 g (81%); mp 175–176°C; [α]_D –24.0 (*c* 1.0, DMF); MS (CI): m/z 464 (5%) [M+H]⁺. ¹H NMR (200 MHz, CDCl₃): δ 7.4–7.2 (m, 5H, Ph), 6.58 (d, 1H, J_{trans} 16.0 Hz, CH₂CH=CHPh), 6.20 (m, 1H, CH₂CH=CHPh), 5.49 (d, 1H, $J_{2,NH}$ 8.8 Hz, NH), 5.28 (t, 1H, $J_{2,3} = J_{3,4}$ 9.3 Hz, H-3), 5.07 (t, 1H, $J_{3,4} = J_{4,5}$ 9.6 Hz, H-4), 4.76 (d, 1H, $J_{1,2}$ 8.3 Hz, H-1), 4.48 (ddd, 1H, J_{gem} 12.8 Hz, J 5.6 Hz, ⁴J 1.4 CH_AH_BCH=CHPh), Hz, 4.3-4.1 3H, (m. CH_AH_BCH=CHPh, H-6, H-6'), 3.87 (m, 1H, H-2), 3.69 (m, 1H, H-5), 2.06, 2.01, 2.00, 1.93 (4s, 4CH₃CO). ¹³C NMR (50 MHz, CDCl₃): δ 170.9, 170.7, 170.2, 169.4 (4C=O), 136.3, 133.3, 128.6, 127.9, 126.5, 124.6 (CH=CHPh), 99.6 (C-1), 72.3 (C-4), 71.8 (C-3), 69.7 (OCH₂R), 68.6 (C-5), 62.1 (C-6), 54.8 (C-2), 23.4 (CH_3CON), 20.7, 20.6, 20.5 ($3CH_3COO$). Anal. calcd for C₂₃H₂₉NO₉: C, 59.60; H, 6.31; N, 3.02. Found: C, 59.37; H, 6.19; N, 2.97%.

3.2.4. 3-Methyl-2-butenyl 2-acetamido-3,4,6-tri-O-acetyl-**2-deoxy-β-D-glucopyranoside**, **5**. Yield 15.1 g (73%); mp 138–139°C; $[\alpha]_D$ +50.0 (c 0.6, CHCl₃); MS (CI): m/z 416 (27%) [M+H]⁺. ¹H NMR (200 MHz, CDCl₃): δ 5.96 (d, 1H, J_{2,NH} 8.8 Hz, NH), 5.3–5.1 [m, 2H, H-3, $CH=C(CH_3)_2$], 4.98 (t, 1H, $J_{3,4}=J_{4,5}$ 9.6 Hz, H-4), 4.65 (d, 1H, J_{1,2} 8.3 Hz, H-1), 4.2–4.0 (m, 4H, H-6, H-6', OCH₂R), 3.78 (m, 1H, H-2), 3.64 (m, 1H, H-5), 2.01, 1.96, 1.95, 1.88 (4s, 4CH₃CO), 1.68, 1.60 [2s, 6H, CH=C(CH₃)₂]. ¹³C NMR (50 MHz, CDCl₃): δ 170.7, 170.6, 170.5, 169.3 (4C=O), 139.0 [CH=<u>C</u>(CH₃)₂], 119.5 [<u>C</u>H=C(CH₃)₂], 98.8 (C-1), 72.3 (C-4), 71.6 (C-3), 68.8 (C-5), 65.0 (OCH₂R), 62.2 (C-6), 54.6 (C-2), 25.7, 23.2, 20.6, 20.5, 20.4, 17.8 (6CH₃). HRMS (CI): [M+H]⁺, found 416.192050. $C_{19}H_{30}NO_9$ requires 416.192057. Anal. calcd for C₁₉H₂₉NO₉: C, 54.93; H, 7.04; N, 3.37. Found: C, 54.93; H, 6.98; N, 3.37%.

3.3. Alkenyl 2-acetamido-(R)-4,6-O-benzylidene-2deoxy- β -D-glucopyranosides, 7–12

To a solution of 1–6 (25 mmol) in methanol (100 mL) was added a solution of sodium methoxide (2.0 mmol) in methanol (20 mL). After 30 min at room temperature, the mixture was neutralized by addition of Dowex 50 resin (H⁺ form), filtered, and evaporated. The solid obtained was dissolved in benzaldehyde (100 mL), and zinc chloride (5 g) was added. The mixture was stirred for 2 days and then poured into hexane/water (1:1) (500 mL) with stirring. The precipitate was filtered off, washed with water and with hexane, and then recrystallized from ethanol.

3.3.1. 2-Methyl-2-propenyl 2-acetamido-(R)-4,6-O-benzylidene-2-deoxy-β-D-glucopyranoside, 8. Yield 6.1 g (67%); mp 262–264°C; $[\alpha]_D$ –50.0 (*c* 1.0, DMF); MS (CI): m/z 364 (85%) [M+H]⁺. ¹H NMR (200 MHz, DMSO- d_6): δ 7.84 (d, 1H, J_{2,NH} 8.2 Hz, NH), 7.5–7.3 (m, 5H, Ph), 5.59 (s, 1H, PhCH), 5.28 (d, 1H, J_{3,OH} 4.9 Hz, OH), 4.92, 4.83 (2bs, 2H, C=CH₂), 4.45 (d, 1H, J_{1,2} 7.8 Hz, H-1), 4.20 (dd, 1H, $J_{5,6e}$ 4.6 Hz, $J_{6e,6a}$ 10.1 Hz, H-6_e), 4.08 (d, 1H, J_{gem} 13.0 Hz, OCH_AH_BR), 3.87 (d, 1H, J_{gem} 13.0 Hz, $OCH_A \underline{H}_B R$), 3.73 (t, 1H, $J_{5,6a} = J_{6e,6a}$ 9.9 Hz, H-6a), 3.6-3.3 (m, 4H, H-2, H-3, H-4, H-5), 1.81 (s, 3H, CH₃CON), 1.64 (s, 3H, CH₃). ¹³C NMR (50 MHz, DMSO-*d*₆): δ 169.0 (C=O), 141.5 (C=CH₂), 137.7, 128.8, 128.0, 126.3 (Ph), 111.8 (C= CH_2), 101.0 (C-1), 100.7 (PhCH), 81.3 (C-4), 71.9 (OCH₂R), 70.3 (C-3), 67.9 (C-6), 65.9 (C-5), 56.1 (C-2), 23.0 (CH₃CON), 19.0 (CH₃). HRMS (CI): [M+H]⁺, found 364.176046. C₁₉H₂₆NO₆ requires 364.176013. Anal. calcd for C₁₉H₂₅NO₆: C, 62.80; H, 6.93; N, 3.85. Found: C, 62.69; H, 6.77; N, 4.06%.

3.3.2. *trans*-2-Butenyl 2-acetamido-(*R*)-4,6-*O*-benzylidene-2-deoxy-β-D-glucopyranoside, 9. Yield 8.5 g (94%); mp 236–238°C; $[\alpha]_D$ –57.7 (*c* 1.0, DMF); MS (CI): *m/z* 364 (45%) [M+H]⁺. ¹H NMR (200 MHz, DMSO-*d*₆): δ 7.82 (d, 1H, *J*_{2,NH} 8.5 Hz, NH), 7.5–7.3 (m, 5H, Ph), 5.7–5.4 (m, 3H, PhCH, CH=CH), 5.28 (d, 1H, *J*_{3,OH} 5.3

Hz, OH), 4.47 (d, 1H, $J_{1,2}$ 8.0 Hz, H-1), 4.3–4.1 (m, 2H, H-6_e, OCH_AH_BR), 3.91 (dd, 1H, *J* 6.0 Hz, J_{gem} 12.3 Hz, OCH_AH_BR), 3.72 (t, 1H, $J_{5,6a} = J_{6e,6a}$ 9.9 Hz, H-6_a), 3.6–3.2 (m, 4H, H-2, H-3, H-4, H-5), 1.80 (s, 3H, CH₃CON), 1.65 (d, 3H, *J* 6.2 Hz, CH₃). ¹³C NMR (50 MHz, DMSO-*d*₆): δ 169.2 (C=O), 137.8, 128.9, 128.1, 126.4 (Ph), 128.2, 127.4 (CH=CH), 100.8 (C-1), 100.7 (PhCH), 81.3 (C-4), 70.4 (C-3), 68.9 (OCH₂R), 67.9 (C-6), 66.0 (C-5), 56.2 (C-2), 23.1 (CH₃CON), 17.6 (CH₃). HRMS (CI): [M+H]⁺, found 364.175819. C₁₉H₂₆NO₆ requires 364.176013. Anal. calcd for C₁₉H₂₅NO₆: C, 62.80; H, 6.93; N, 3.85. Found: C, 62.63; H, 6.87; N, 3.88%.

3.3.3. trans-3-Phenyl-2-propenyl 2-acetamido-(R)-4,6-Obenzylidene-2-deoxy-β-D-glucopyranoside, 10. Yield 9.2 g (87%); mp 252–253°C; $[\alpha]_D$ –90.0 (*c* 1.2, DMF); MS (FAB): *m*/*z* 448 (100%) [M+Na]⁺. ¹H NMR (200 MHz, DMSO-d₆): δ 7.90 (d, 1H, J_{2,NH} 8.4 Hz, NH), 7.5–7.2 (m, 10H, 2Ph), 6.59 (d, 1H, J_{trans} 16.0 Hz, CH₂CH=C<u>H</u>), 6.27 (dt, 1H, J 5.4 Hz, J_{trans} 16.0 Hz, CH₂CH=CH), 5.60 (s, 1H, PhCH), 5.32 (d, 1H, J_{3,OH} 5.2 Hz, OH), 4.56 (d, 1H, $J_{1,2}$ 7.9 Hz, H-1), 4.37 (ddd, 1H, J_{gem} 12.3 Hz, J 6.0 Hz, ⁴J 1.3 Hz, $OCH_AH_BCH=CH),$ 4.2-4.1 (m, 2H. H-6_e, OCH_AH_BCH=CH), 3.75 (t, 1H, J_{5,6a}=J_{6e,6a} 9.9 Hz, H-6_a), 3.6–3.3 (m, 4H, H-2, H-3, H-4, H-5), 1.83 (s, 3H, CH₃CON). ¹³C NMR (50 MHz, DMSO- d_6): δ 169.2 (C=O), 137.7, 131.0, 128.9, 128.7, 128.0, 127.6, 126.3, 126.0 (Ph, CH=CHPh), 101.0 (C-1), 100.7 (PhCH), 81.3 (C-4), 70.4 (C-3), 68.7 (OCH₂R), 67.9 (C-6), 66.0 (C-5), 56.2 (C-2), 23.2 (CH₃CON), 17.6 (CH₃). HRMS (CI): $[M+H]^+$, found 426.191285. $C_{24}H_{28}NO_6$ requires 426.191663. Anal. calcd for C₂₄H₂₇NO₆: C, 67.75; H, 6.40; N, 3.29. Found: C, 67.53; H, 6.41; N, 3.16%.

3.3.4. 3-Methyl-2-butenyl 2-acetamido-(R)-4,6-O-benzylidene-2-deoxy-β-D-glucopyranoside, 11. Yield 7.1 g (75%); mp 164–166°C; $[\alpha]_D$ –34.6 (*c* 1.0, DMF); MS (CI): m/z 378 (20%) [M+H]⁺. ¹H NMR (200 MHz, DMSO- d_6): δ 7.80 (d, 1H, $J_{2,NH}$ 8.6 Hz, NH), 7.5–7.3 (m, 5H, Ph), 5.58 (s, 1H, PhCH), 5.3-5.1 [m, 2H, OH, CH=C(CH₃)₂], 4.49 (d, 1H, J_{1,2} 8.1 Hz, H-1), 4.2–3.9 (m, 3H, H-6_e, OCH₂R), 3.75 (t, 1H, $J_{5,6a} = J_{6e,6a}$ 9.9 Hz, H-6,), 3.6–3.3 (m, 4H, H-2, H-3, H-4, H-5), 1.81 (s, 3H, CH₃CON), 1.69, 1.59 [2s, 6H, CH=C(CH₃)₂]. ¹³C NMR (50 MHz, DMSO-d₆): δ 169.1 (C=O), 137.8, 128.9, 128.0, 126.4 (Ph), 136.6 [CH=C(CH₃)₂], 120.5 [CH=<u>C</u>(CH₃)₂], 100.7 (Ph<u>C</u>H), 100.5 (C-1), 81.3 (C-4), 70.5 (C-3), 67.9 (OCH₂R), 66.0 (C-5), 64.6 (C-6), 56.2 (C-2), 23.1 (CH_3CON), 25.5, 19.0 [$CH=C(CH_3)_2$]. HRMS (CI): [M+H]⁺, found 378.191480. C₂₀H₂₈NO₆ requires 378.191663. Anal. calcd for C₂₀H₂₇NO₆: C, 63.65; H, 7.21; N, 3.71. Found: C, 63.39; H, 6.99; N, 3.97%.

3.3.5. 3-Butenyl 2-acetamido-(*R*)-**4**,**6**-*O*-benzylidene-2deoxy-β-D-glucopyranoside, **12**. Yield 6.0 g (66%); mp 235–236°C; [α]_D –44.4 (*c* 0.9, DMF); MS (CI): *m*/*z* 364 (100%) [M+H]⁺. ¹H NMR (200 MHz, CDCl₃): δ 7.5–7.3 (m, 5H, Ph), 6.0–5.7 (m, 2H, NH, CH=CH₂), 5.51 (s, 1H, PhCH), 5.08 (m, 2H, CH=CH₂), 4.64 (d, 1H, *J*_{1,2} 8.2 Hz, H-1), 3.6–3.4 (m, 4H, H-2, H-3, H-4, H-5), 2.34 (m, 2H, OCH₂CH₂R), 2.02 (s, 3H, CH₃CON). ¹³C NMR (50 MHz, CDCl₃): δ 172.2 (C=O), 137.0, 129.2, 128.3, 126.3 (Ph), 134.9 (CH=CH₂), 117.0 (CH=CH₂), 101.9 (C-1), 100.5 (PhCH), 81.5 (C-4), 71.2 (C-3), 68.8 (OCH₂CH₂R), 68.5 (C-6), 66.3 (C-5), 59.2 (C-2), 33.8 (OCH₂CH₂R), 23.6 (CH₃CON). HRMS (CI): [M+H]⁺, found 364.175683. C₁₉H₂₆NO₆ requires 364.176013. Anal. calcd for C₁₉H₂₅NO₆: C, 62.80; H, 6.93; N, 3.85. Found: C, 62.73; H, 6.93; N, 3.86%.

3.4. Allyl 2-acetamido-3-*O*-acetyl-(*R*)-4,6-*O*-benzylidene-2-deoxy-β-D-glucopyranosides, 13

Allyl 2-acetamido-(R)-4,6-O-benzylidene-2-deoxy- β -Dglucopyranoside 7 (3.5 g, 10 mmol) was acetylated in the usual way with acetic anhydride/pyridine (1:1) (40 mL). The reaction mixture was kept overnight at room temperature and then poured into ice-water. The precipitate obtained was isolated by filtration and, recrystallized from ethanol, yielded 3.7 g (95%); mp 283–284°C; $[\alpha]_D$ –77.1 (c 0.7, CH₂Cl₂); MS (CI): m/z392 (60%) $[M+H]^+$. ¹H NMR (200 MHz, CDCl₃): δ 7.5–7.3 (m, 5H, Ph), 5.98 (d, 1H, J_{2,NH} 9.5 Hz, NH), 5.77 (m, 1H, CH=CH₂), 5.48 (s, 1H, PhCH), 5.3–5.1 (m, 3H, H-3, CH=C \underline{H}_2), 4.44 (d, 1H, $J_{1,2}$ 8.4 Hz, H-1), 4.3-4.05 (m, 3H, H-2, H-6e, OCHAHBR), 3.89 (m, 1H, OCH_AH_BR), 3.8–3.65 (m, 2H, H-4, H-6_a), 3.52 (dt, 1H, $J_{4.5} = J_{5.6a}$ 9.6 Hz, $J_{5.6e}$ 4.7 Hz, H-5), 2.07 (s, 3H, CH₃COO), 1.95 (s, 3H, CH₃CON). ¹³C NMR (50 MHz, CDCl₃): δ 171.4, 170.2 (2C=O), 136.9, 129.1, 128.2, 126.0 (Ph), 133.5 (CH=CH₂), 117.4 (CH=CH₂), 101.2 (C-1), 101.1 (PhCH), 78.6 (C-4), 72.0 (C-3), 70.0 (OCH₂R), 68.6 (C-6), 66.2 (C-5), 54.3 (C-2), 23.3 (CH₃CON), 20.9 (CH₃COO). HRMS (CI): [M+H]⁺, found 392.170795. C₂₀H₂₆NO₇ requires 392.170928. Anal. calcd for C₂₀H₂₅NO₇: C, 61.37; H, 6.44; N, 3.58. Found: C, 61.21; H, 6.35; N, 3.62%.

3.5. Alkenyl 2-acetamido-3-*O*-alkyl-(*R*)-4,6-*O*-benzylidene-2-deoxy-β-D-glucopyranosides, 14 and 15

3.5.1. Allyl 2-acetamido-(R)-4,6-O-benzylidene-2-deoxy-**3-***O***-methyl-β-D-glucopyranoside**, **14**. To a solution of 7 (3.5 g, 10 mmol) in dry THF (50 mL) were added, successively, freshly powdered potassium hydroxide (1.0 g, 17.9 mmol), 18-crown-6 (150 mg, 0.5 mmol), and methyl iodide (20.8 mmol). The mixture was stirred at room temperature, and the reaction was monitored by TLC at the end of the reaction. The mixture was then diluted with dichloromethane, and washed several time with water and dried (Na_2SO_4) . Evaporation of the organic phase gave the pure methylated product. The solid obtained, purified by crystallization from ethanol, yielded 2.3 g (64%); mp 290–291°C; [α]_D –103.4 (c 0.7, DMF); MS (CI): m/z 364 (100%) [M+H]⁺. ¹H NMR (200 MHz, DMSO-d₆): δ 7.96 (d, 1H, J_{2.NH} 8.7 Hz, NH), 7.5–7.3 (m, 5H, Ph), 5.82 (m, 1H, CH=CH₂), 5.65 (s, 1H, PhCH), 5.15 (m, 2H, CH=CH₂), 4.52 (d, 1H, $J_{1,2}$) 8.0 Hz, H-1), 4.3–4.1 (m, 2H, H-6_e, $OC\underline{H}_{A}H_{B}R$), 4.00 (m, 1H, OCH_A $\underline{H}_{B}R$), 3.77 (t, 1H, $J_{5,6a} = J_{6e,6a}$ 10.1 Hz, H-6_a), 3.6–3.4 (m, 4H, H-2, H-3, H-4, H-5), 3.35 (s, 3H, OCH₃), 1.82 (s, 3H, CH₃CON). ¹³C NMR (50 MHz, DMSO- d_6): δ 169.0 (C=O), 137.6, 128.8, 128.1, 126.0 (Ph), 134.5 (CH=CH₂), 116.2 (CH=CH₂), 101.0 (C-1), 100.2 (PhCH), 80.6 (C-4), 79.5 (C-3), 69.0 (OCH₂R), 67.8 (C-6), 65.6 (C-5), 59.0 (OCH₃), 54.2 (C-2), 22.9 (CH₃CON). HRMS (CI): $[M+H]^+$, found 364.175979. C₁₉H₂₆NO₆ requires 364.176013. Anal. calcd for C₁₉H₂₅NO₆: C, 62.80; H, 6.93; N, 3.85. Found: C, 62.49; H, 6.70; N, 3.85%.

3.5.2. Allyl 2-acetamido-3-O-benzyl-(R)-4,6-O-benzylidene-2-deoxy-β-D-glucopyranoside, 15. To a solution of 7 (3.5 g, 10 mmol) in distilled dichloromethane (200 mL), benzyl bromide (2.5 mL, 21.0 mmol), 50% aqueous sodium hydroxide solution (200 mL) and tetrabutylammonium bromide (32 mg, 0.10 mmol) were added. The reaction mixture was stirred vigorously at room temperature for 3 h. The organic phase was washed with water until neutral, dried (MgSO₄), and the solvent was removed under reduced pressure to give a solid which, purified by crystallization from ethyl acetate-ethanol, yielded 3.1 g (71%); mp 270-272°C; $[\alpha]_{\rm D}$ -37.6 (c 1.0, DMF); MS (CI): m/z 440 (10%) $[M+H]^+$. ¹H NMR (200 MHz, DMSO-*d*₆): δ 8.00 (d, 1H, J_{2,NH} 8.4 Hz, NH), 7.5–7.2 (m, 10H, 2Ph), 5.81 (m, 1H, $CH=CH_2$), 5.70 (s, 1H, PhCH), 5.18 (m, 2H, CH=CH_2), 4.72 (d, 1H, J_{gem} 11.9 Hz, PhCH_AH_BO), 4.6–4.5 (m, 2H, PhCH_AH_BO, H-1), 4.3–4.15 (m, 2H, $H-6_{e}$, $OCH_{A}H_{B}R$), 4.01 (m, 1H, $OCH_{A}H_{B}R$), 3.8–3.6 (m, 4H, H-2, H-3, H-4, H-6_a), 3.46 (m, 1H, H-5), 1.81 (s, 3H, CH₃CON). ¹³C NMR (50 MHz, DMSO- d_6): δ 169.1 (C=O), 138.7, 137.6, 129.7, 128.8, 128.1, 128.0, 127.4, 126.0 (2Ph), 134.5 (CH=CH₂), 116.3 (CH=CH₂), 100.9 (C-1), 100.1 (PhCH), 80.9 (C-4), 78.6 (C-3), 73.2 (PhCH2O), 69.0 (OCH2R), 67.8 (C-6), 65.6 (C-5), 54.6 (C-2), 23.0 (CH₃CON). HRMS (CI): [M+H]⁺, found 440.207175. C₂₅H₃₀NO₆ requires 440.207313. Anal. calcd for C₂₅H₂₉NO₆: C, 68.32; H, 6.65; N, 3.19. Found: C, 68.15; H, 6.97; N, 3.34%.

3.6. Alkenyl 2-acetamido-(R)-4,6-O-benzylidene-2deoxy-3-O-tert-butyldimethylsilyl- β -D-glucopyranosides, 16 and 17

To a solution of 7, 8 (10 mmol) in dry DMF (30 mL), imidazole (3.4 g, 50 mmol) and *tert*-butyldimethylsilyl chloride (3.8 g, 25 mmol) were added and the mixture was stirred at room temperature for 2 h. The reaction mixture was diluted with dichloromethane, washed with water, dried (Na₂SO₄), filtered and the filtrate evaporated to dryness to give a white solid, which was purified by flash chromatography on silica gel using hexane–ethyl acetate (7:1).

3.6.1. Allyl 2-acetamido-(R)-4,6-O-benzylidene-2-deoxy-3-O-tert-butyldimethylsilyl-β-D-glucopyranoside, 16. Yield 3.8 g (82%); mp 98–99°C; $[\alpha]_D$ -66.3 (c 1.0, CH₂Cl₂); MS (CI): m/z 464 (60%) [M+H]⁺. ¹H NMR (200 MHz, DMSO- d_6): δ 7.91 (d, 1H, $J_{2.NH}$ 9.0 Hz, NH), 7.5–7.3 (m, 5H, Ph), 5.82 (m, 1H, CH=CH₂), 5.63 (s, 1H, PhCH), 5.17 (m, 2H, CH=CH₂), 4.50 (d, 1H, $J_{1,2}$) 8.3 Hz, H-1), 4.3-4.1 (m, 2H, H-6e, OCHAHBR), 3.99 (m, 1H, OCH_AH_BR), 3.8–3.4 (m, 5H, H-2, H-3, H-4, H-5, H-6_a), 1.78 (s, 3H, CH₃CON), 0.75 [s, 9H, $OSi(CH_3)_2C(CH_3)_3],$ -0.03, -0.106H, [2s,

OSi(CH₃)₂C(CH₃)₃]. ¹³C NMR (50 MHz, DMSO-*d*₆): δ 168.8 (C=O), 137.6, 128.8, 128.0, 126.2 (Ph), 134.6 (CH=CH₂), 116.2 (CH=CH₂), 100.8 (C-1), 100.7 (PhCH), 81.5 (C-4), 72.3 (C-3), 68.9 (OCH₂R), 67.8 (C-6), 65.6 (C-5), 56.0 (C-2), 25.6 [OSi(CH₃)₂C(CH₃)₃], 23.1 (CH₃CON), 17.9 [OSi(CH₃)₂C(CH₃)₃], -4.2, -5.0 [OSi(CH₃)₂C(CH₃)₃]. HRMS (CI): [M+H]⁺, found 464.246313. C₂₄H₃₈NO₆Si requires 464.246842. Anal. calcd for C₂₄H₃₇NO₆Si: C, 62.17; H, 8.04; N, 3.02. Found: C, 62.11; H, 7.90; N, 3.15%.

3.6.2. 2-Methyl-2-propenyl 2-acetamido-(R)-4,6-O-benzylidene-2-deoxy-3-O-tert-butyldimethylsilyl-β-D-gluco**pyranoside**, 17. Yield 3.4 g (71%); mp 132–133°C; $[\alpha]_{D}$ -30.4 (c 0.9, CH₂Cl₂); MS (CI): m/z 478 (75%) [M+H]⁺. ¹H NMR (200 MHz, CDCl₃): δ 7.5–7.3 (m, 5H, Ph), 5.87 (d, 1H, J_{2,NH} 8.1 Hz, NH), 5.46 (s, 1H, PhCH), 4.90 (m, 3H, C=CH₂, H-1), 4.3-4.2 (m, 2H, H-6_e, OC $\underline{H}_A H_B R$), 3.95 (d, 1H, J_{gem} 12.6 Hz, OC $\underline{H}_A \underline{H}_B R$), 3.73 (t, 1H, $J_{5,6a} = J_{6e,6a}$ 10.0 Hz, H-6_a), 3.5–3.3 (m, 4H, H-2, H-3, H-4, H-5), 1.95 (s, 3H, CH₃CON), 1.69 (s, 3H, CH₃), 0.79 [s, 9H, OSi(CH₃)₂C(CH₃)₃], -0.02, -0.08 [2s, 6H, OSi(CH₃)₂C(CH₃)₃]. ¹³C NMR (50 MHz, CDCl₃): δ 170.0 (C=O), 141.1 (C=CH₂), 137.2, 128.9, 128.1, 126.2 (Ph), 112.8 (C=CH₂), 101.7 (C-1), 99.5 (PhCH), 82.5 (C-4), 73.0 (OCH₂R), 71.1 (C-3), 68.7 (C-6), 65.8 (C-5), 59.2 (C-2), 25.7 [OSi(CH₃)₂C(CH₃)₃], 23.7 (CH₃CON), 19.3 (CH₃), 18.1 [OSi(CH₃)₂C(CH₃)₃], -4.2, -5.0 [OSi(CH₃)₂C(CH₃)₃]. HRMS (CI): [M+H]⁺, found 478.261653. C₂₅H₄₀NO₆Si requires 478.262492. Anal. calcd for C₂₅H₃₉NO₆Si: C, 62.86; H, 8.23; N, 2.93. Found: C, 62.79; H, 8.18; N, 3.01%.

3.7. Allyl 2-acetamido-(R)-4,6-O-benzylidene-2-deoxy-2-N-3-O-methylidene- β -D-glucopyranoside, 18

To a solution of allyl 2-acetamido-(R)-4,6-O-benzylidene-2-deoxy-β-D-glucopyranoside 7 (10 mmol) in distilled dichloromethane (50 mL), dibromomethane (50 mL), 50% aqueous sodium hydroxide solution (100 mL), and tetrabutylammonium bromide (6.5 mg, 0.02 mmol) were added. The reaction mixture was vigorously stirred under reflux for 2 days, then cooled to room temperature. The organic phase was washed with water until neutral, dried (MgSO₄), and the solvent was removed under reduced pressure to give a solid which was purified by flash chromatography on silica gel using hexane-ethyl acetate (15:10) as eluent, yielding 3.5 g (97%); mp 94–95°C; $[\alpha]_D$ –21.5 (*c* 1.0, CH₂Cl₂); MS (CI): *m*/*z* 362 (100%) [M+H]⁺. ¹H NMR (200 MHz, CDCl₃): δ 7.96 (d, 1H, J_{2,NH} 8.7 Hz, NH), 7.5–7.3 (m, 5H, Ph), 5.90 (m, 1H, CH=CH₂), 5.60 (s, 1H, PhCH), 5.48 (d, 1H, J_{gem} 5.3 Hz, OCH_AH_BN), 5.28 (m, 2H, CH=CH₂), 4.8-4.7 (m, 2H, OCH_AH_BN, H-1), 4.5-4.3 (m, 2H, OCH_AH_BR , H-6_e), 4.14 (m, 1H, OCH_AH_BR), 4.0-3.8 (m, 3H, H-3, H-4, H-6_a), 3.50 (m, 1H, H-5), 3.37 (dd, 1H, J_{1,2} 7.9 Hz, J_{2,3} 9.2 Hz, H-2), 2.22 (s, 3H, CH₃CON). ¹³C NMR (50 MHz, CDCl₃): δ 173.3 (C=O), 136.5, 129.3, 128.3, 126.2 (Ph), 132.8 (CH=CH₂), 118.7 (CH=CH₂), 102.8 (C-1), 101.6 (PhCH), 82.4 (OCH2N), 82.0 (C-4), 79.2 (C-3), 70.6 (OCH₂R), 69.1 (C-5), 68.5 (C-6), 62.0 (C-2), 23.2 (CH₃CON). Anal. calcd for $C_{19}H_{23}NO_6$: C, 63.15; H, 6.41; N, 3.88. Found: C, 62.88; H, 6.55; N, 3.82%.

3.8. Epoxidation of alkenyl glucosides with *m*-chloroperoxybenzoic acid

To a solution of either 1, 2, 4, 5 or 7–18 (1.5 mmol) in distilled dichloromethane (300 mL), *m*-chloroperoxybenzoic acid (Aldrich 57–86%) (3.0 g) was added and the suspension was stirred. When TLC showed that all starting compound had been consumed, the reaction mixture was washed successively with a 5% aqueous solution of sodium hydroxide and water, dried (MgSO₄), filtered, and the filtrate evaporated to dryness. Diastereomeric excess (d.e.) was determined by ¹H NMR. The solids obtained were purified by recrystallization from ethanol or by flash chromatography on silica gel.

3.8.1. Epoxidation of peracetyl derivatives

3.8.1.1. (R)-2,3-Epoxy-2-methylpropyl 2-acetamido-**3,4,6-tri-***O***-acetyl-2-deoxy-**β**-**D**-glucopyranoside**, **20**. Mp 152–154°C; $[\alpha]_{D}$ +16.6 (*c* 0.8, DMF); MS (CI): *m*/*z* 418 (15%) [M+H]⁺. ¹H NMR (200 MHz, CDCl₃): δ 5.64 (d, 1H, J_{2,NH} 8.8 Hz, NH major), 5.53 (d, J_{2,NH} 8.6 Hz, NH minor), 4.67 (d, J_{1,2} 8.4 Hz, H-1 minor), 4.54 (d, J_{1.2} 8.5 Hz, H-1 major), 2.82, 2.55 [2d, 2H, J_{gem} 5.1 Hz, OCH₂C(CH₃)(O)CH₂], 2.06, 2.01, 2.00, 1.99, 1.94 (4s, 12H, 4CH₃CO), 1.33 (s, CH₃ minor), 1.29 (s, CH₃ major). ¹³C NMR (50 MHz, CDCl₃): δ 170.9, 170.7, 170.3, 170.2, 169.3 (4C=O), 101.6 (C-1 major), 101.2 (C-1 minor) 72.7, 72.5, 71.9, 71.8 (C-4, C-3), 69.6 (C-5 minor), 68.4 (C-5 major), 70.6 [OCH₂C(CH₃)(O)CH₂ minor], 69.8 [OCH2C(CH3)(O)CH2 major], 62.0 (C-6 major), 61.5 (C-6 minor), 56.1 [OCH₂C(CH₃)(O)CH₂ minor], 56.0 [OCH2C(CH3)(O)CH2 major], 54.2 (C-2 major), 51.0 (C-2 minor), 51.6 [OCH₂C(CH₃)(O)CH₂ minor], 50.4 [OCH₂C(CH₃)(O)CH₂ major], 23.2, 23.0, 20.9, 20.7, 20.6, 20.5, 18.3, 18.1 (5CH₃). HRMS (CI): $[M+H]^+$, found 418.171028. $C_{18}H_{28}NO_{10}$ requires 418.171322. Anal. calcd for $C_{18}H_{27}NO_{10}$: C, 51.79; H, 6.52; N, 3.36. Found: C, 51.52; H, 6.33; N, 3.48%.

3.8.1.2. (2S,3S)-2,3-Epoxy-3-phenylpropyl 2-acetamido-3,4,6-tri-O-acetyl-2-deoxy-β-D-glucopyranoside, 21. Mp 141–142°C; $[\alpha]_D$ –32.0 (*c* 1.0, DMF); MS (CI): *m*/*z* 480 (35%) [M+H]⁺. ¹H NMR (200 MHz, CDCl₃): δ 7.4-7.2 (m, 5H, Ph), 5.78 (d, 1H, J_{2,NH} 8.9 Hz, NH), 5.18 (t, 1H, $J_{2,3} = J_{3,4}$ 9.5 Hz, H-3), 5.06 (t, 1H, $J_{3,4} = J_{4,5}$ 9.4 Hz, H-4), 4.81 (d, $J_{1,2}$ 8.5 Hz, H-1 minor), 4.65 (d, $J_{1,2}$ 8.4 Hz, H-1 major), 4.25 [dd, 1H, J_{gem} 12.2 Hz, J 4.6 Hz, $OCH_AH_BCH(O)CHPh$], 4.14 [dd, 1H, J_{gem} 12.5 Hz, J 2.2 Hz, OCH_AH_BCH(O)CHPh], 4.05–3.95 (m, 3H, H-2, H-6, H-6'), 3.85 [d, 1H, J 2.0 Hz, OCH₂CH(O)CHPh], 3.68 (m, 1H, H-5), 3.17 [m, 1H, OCH₂CH(O)CHPh], 2.07, 2.02, 2.01, 1.93 (4s, 4CH₃CO). ¹³C NMR (50 MHz, CDCl₃): δ 170.9, 170.7, 170.3, 169.3 (4C=O), 136.4, 134.5, 128.5, 128.4, 125.6 (Ph), 101.6 (C-1 major), 100.8 (C-1 minor), 72.5 (C-4), 71.9 (C-3), 68.3 (C-5), 67.0 [OCH₂CH(O)CHPh], 62.0 (C-6), 60.8 [OCH₂CH(O)CHPh], 55.1 (C-2), 54.3 [OCH₂CH(O)CHPh], 23.3 (CH₃CON), 20.8, 20.7, 20.6 (3CH₃COO). HRMS (EI): [M]+[•], found 479.177804.

 $C_{23}H_{29}NO_{10}$ requires 479.179147. Anal. calcd for $C_{23}H_{29}NO_{10}$: C, 57.61; H, 6.10; N, 2.92. Found: C, 57.38; H, 6.00; N, 3.10%.

3.8.1.3. (S)-2,3-Epoxy-3-methylbutyl 2-acetamido-3,4,6-tri-O-acetyl-2-deoxy-β-D-glucopyranoside, 22. Mp 156–158°C; $[\alpha]_{D}$ +13.9 (*c* 0.9, DMF); MS (CI): *m*/*z* 432 (65%) $[M+H]^+$. ¹H NMR (200 MHz, CDCl₃): δ 5.82 (2d, 1H, J_{2.NH} 8.4 Hz, 9.0 Hz, NH), 4.79 (d, J_{1.2} 8.3 Hz, H-1 minor), 4.70 (d, J_{1,2} 8.4 Hz, H-1 major), 2.92 [m, 1H, OCH₂CH(O)C(CH₃)₂], 2.04, 1.99, 1.98, 1.92, 1.90 (4s, 12H, 4CH₃CO), 1.28, 1.24 [2s, 6H, OCH₂CH(O)-C(CH₃)₂]. ¹³C NMR (50 MHz, CDCl₃): δ 170.9, 170.7, 170.3, 169.4, 169.3 (4C=O), 101.8 (C-1 major), 100.2 (C-1 minor) 72.5, 72.1, 71.9, 71.8 (C-4, C-3), 68.6 (C-5 minor), 68.4 (C-5 major), 68.3 [OCH₂CH(O)C(CH₃)₂ major], 67.9 [OCH₂CH(O)C(CH₃)₂ minor], 62.1 (C-6 minor), 62.0 (C-6 major), 62.2 [OCH₂CH(O)C(CH₃)₂ $[OCH_2CH(O)C(CH_3)_2 \text{ minor}],$ major], 61.1 58.2 [OCH₂CH(O)C(CH₃)₂ minor], 57.7 [OCH₂CH(O)C-(CH₃)₂ major], 54.8 (C-2 minor), 54.3 (C-2 major), 24.6, 24.5, 23.3, 20.7, 20.6, 18.9, 18.7 (6CH₃). HRMS (CI): $[M+H]^+$, found 432.185043. $C_{19}H_{30}NO_{10}$ requires 432.186972. Anal. calcd for $C_{19}H_{29}NO_{10}$: C, 52.89; H, 6.78; N, 3.25. Found: C, 52.66; H, 6.57; N, 3.39%.

3.8.2. Epoxidation of 4,6-O-benzylidene derivatives

3.8.2.1. (R)-2,3-Epoxy-2-methylpropyl 2-acetamido-(R)-4,6-O-benzylidene-2-deoxy- β -D-glucopyranoside, 24. Mp 274–276°C; [α]_D –69.1 (*c* 1.0, DMF); MS (CI): *m*/*z* 380 (100%) $[M+H]^+$. ¹H NMR (200 MHz, CDCl₃): δ 7.5–7.3 (m, 5H, Ph), 6.85 (d, 1H, J_{2,NH} 3.8 Hz, NH), 5.54 (s, 1H, PhCH), 4.67 (d, J_{1,2} 8.4 Hz, H-1 minor), 4.44 (d, J_{1.2} 8.4 Hz, H-1 major), 4.31 (dd, 1H, J_{5.6e} 4.9 Hz, $J_{6e,6a}$ 10.3 Hz, H-6_e), 3.40 (m, 1H, H-5), 2.95, 2.66 [2d, 2H, J_{gem} 4.9 Hz, OCH₂C(CH₃)(O)CH₂], 2.05 (s, 3H, CH₃CON), 1.39 (s, CH₃ minor), 1.34 (s, CH₃ major). ¹³C NMR (50 MHz, CDCl₃): δ 173.1 (C=O), 137.0, 129.1, 128.2, 126.3 (Ph), 102.9 (C-1 major), 102.8 (C-1 minor), 101.9 (PhCH), 81.3 (C-4), 73.3 (C-3), 69.5 [OCH₂C(CH₃)(O)CH₂], 68.4 (C-6), 66.3 (C-5), 59.6 (C-2), 57.1 [OCH₂C(CH₃)(O)CH₂], 49.6 [OCH₂C(CH₃)(O)- CH_2], 23.0 (CH₃CON), 18.2 [OCH₂C(CH₃)(O)CH₂]. HRMS (CI): [M+H]⁺, found 380.171042. C₁₉H₂₆NO₇ requires 380.170928. Anal. calcd for C19H25NO7: C, 60.15; H, 6.64; N, 3.69. Found: C, 60.10; H, 6.70; N, 3.71%.

3.8.2.2. (2*S*,3*S*)-2,3-Epoxybutyl 2-acetamido-(*R*)-4,6-*O*-benzylidene-2-deoxy-β-D-glucopyranoside, 25. Mp 258–260°C; $[\alpha]_D$ –76.7 (*c* 0.7, DMF); MS (CI): *m*/*z* 380 (100%) [M+H]⁺. ¹H NMR (200 MHz, CDCl₃): δ 7.5– 7.3 (m, 5H, Ph), 6.70 (d, 1H, $J_{2,NH}$ 4.8 Hz, NH), 5.54 (s, 1H, PhCH), 4.68 (d, $J_{1,2}$ 8.3 Hz, H-1 minor), 4.49 (d, $J_{1,2}$ 8.4 Hz, H-1 major), 4.31 (dd, 1H, $J_{5,6e}$ 4.9 Hz, $J_{6e,6a}$ 10.4 Hz, H-6_e), 4.08 [dd, 1H, J_{gem} 13.3 Hz, J 2.1 Hz, OCH_AH_BCH(O)CHCH₃], 3.74 (t, 1H, $J_{5,6a}=J_{6e,6a}$ 10.0 Hz, H-6_a), 3.40 (m, 1H, H-5), 3.10 [m, 1H, OCH₂CH(O)CHCH₃], 2.89 [m, 1H, OCH₂CH(O)-CHCH₃], 2.05 (s, 3H, CH₃CON), 1.40 [d, J 5.3 Hz, OCH₂CH(O)CHCH₄ major]. ¹³C NMR (50 MHz, DMSO-*d*₆): δ 169.2 (C=O), 137.7, 128.9, 128.0, 126.4 (Ph), 101.7 (C-1 major), 101.6 (C-1 minor), 100.7 (PhCH), 81.2 (C-4), 70.3 (C-3), 69.0 [OCH₂CH(O)-CHCH₃ minor], 68.8 [OCH₂CH(O)CHCH₃ major], 67.8 (C-6), 66.0 (C-5), 56.9 [OCH₂CH(O)CHCH₃ minor], 56.7 [OCH₂CH(O)CHCH₃ major], 56.1 (C-2), 51.3 [OCH₂CH(O)CHCH₃ minor], 51.2 [OCH₂CH(O)-CHCH₃ major], 23.1 (CH₃CON), 17.1 [OCH₂CH(O)-CHCH₃]. HRMS (CI): [M+H]⁺, found 380.171278. C₁₉H₂₆NO₇ requires 380.170928. Anal. calcd for C₁₉H₂₅NO₇: C, 60.15; H, 6.64; N, 3.69. Found: C, 59.88; H, 6.64; N, 3.82%.

3.8.2.3. (2S,3S)-2,3-Epoxy-3-phenylpropyl 2-acetamido-(*R*)-4,6-*O*-benzylidene-2-deoxy-β-D-glucopyranoside, **26**. Mp 248–250°C; $[\alpha]_D$ –88.3 (*c* 1.2, DMF); MS (FAB): *m*/*z* 464 (100%) [M+Na]⁺. ¹H NMR (500 MHz, DMSO-*d*₆): δ 7.84 (d, 1H, *J*_{2,NH} 8.8 Hz, NH), 7.5–7.2 (m, 10H, 2Ph), 5.59 (s, 1H, PhCH), 5.28 (d, 1H, J_{3,OH} 5.5 Hz, OH), 4.59 (d, J_{1.2} 8.2 Hz, H-1 minor), 4.55 (d, $J_{1,2}$ 8.2 Hz, H-1 major), 4.19 (dd, $J_{5,6e}$ 5.0 Hz, $J_{6e,6a}$ 10.2 Hz, H-6_e major), 4.11 (dd, $J_{5,6e}$ 4.9 Hz, $J_{6e,6a}$ 10.2 Hz, H-6_e minor), 3.85 [m, 2H, OCH_AH_BCH(O)CHPh], 3.78 [dd, 1H, J_{gem} 12.3 Hz, J 5.0 Hz, OCH_AH_BCH(O)-CHPh], 3.73 (t, 1H, $J_{5,6a} = J_{6e,6a}$ 10.0 Hz, H-6_a), 3.62 (m, 1H, H-3), 3.56 (m, 1H, H-2), 3.44 (t, 1H, $J_{3,4}=J_{4,5}$ 9.6 Hz, H-4), 3.35 (dt, 1H, $J_{4,5}=J_{5,6a}$ 9.8 Hz, $J_{5,6e}$ 4.9 Hz, H-5), 3.24 [m, 1H, OCH₂CH(O)CHPh], 1.82 (s, 3H, CH₃CON major), 1.80 (s, 3H, CH₃CON minor). ¹³C NMR (125 MHz, DMSO- d_6): δ 169.3 (C=O), 137.7, 137.0, 128.9, 128.5, 128.2, 128.1, 126.4, 125.9 (2Ph), 102.0 (C-1 major), 101.9 (C-1 minor), 100.7 (PhCH), 81.2 (C-4), 70.4 (C-3), 68.8 [OCH2CH(O)CHPh minor], 68.3 [OCH₂CH(O)CHPh major], 67.8 (C-6), 66.0 (C-5), 60.3 [OCH2CH(O)CHPh minor], 60.1 [OCH2CH(O)-CHPh major], 56.1 (C-2), 55.3 [OCH₂CH(O)CHPh 54.9 [OCH₂CH(O)CHPh major], minor], 23.1 (CH₃CON). HRMS (CI): [M+H]⁺, found 442.185556. C₂₄H₂₈NO₇ requires 442.186578. Anal. calcd for C₂₄H₂₇NO₇: C, 65.29; H, 6.16; N, 3.17. Found: C, 65.13; H, 6.11; N, 3.11%.

3.8.2.4. (S)-2,3-Epoxy-3-methylbutyl 2-acetamido-(R)-**4,6-***O*-benzylidene-2-deoxy-β-D-glucopyranoside, 27. Mp 188–190°C; [α]_D –64.9 (*c* 1.0, DMF); MS (CI): *m*/*z* 394 (80%) [M+H]⁺. ¹H NMR (200 MHz, CDCl₃): δ 7.5–7.3 (m, 5H, Ph), 6.35 (d, J_{2,NH} 5.7 Hz, NH minor), 6.18 (d, J_{2.NH} 5.8 Hz, NH major), 5.53 (s, 1H, PhCH), 4.75 (d, $J_{1,2}$ 8.3 Hz, H-1 minor), 4.67 (d, $J_{1,2}$ 8.3 Hz, H-1 major), 4.30 (dd, 1H, $J_{5,6e}$ 4.9 Hz, $J_{6e,6a}$ 10.3 Hz, H-6), 3.76 (t, 1H, $J_{5,6a} = J_{6e,6a}$ 10.1 Hz, H-6_a), 2.99 [dd, 1H, J 3.5 Hz, 7.9 Hz, OCH₂CH(O)C(CH₃)₂], 2.04 (s, CH₃CON major), 2.02 (s, CH₃CON minor), 1.34, 1.31 [2s, OCH₂CH(O)C(CH₃)₂ minor], 1.33, 1.29 [2s, OCH₂CH(O)C(CH₃)₂ major]. ¹³C NMR (50 MHz, DMSO- d_6): δ 169.2 (C=O), 137.7, 128.8, 128.0, 126.3 (Ph), 101.7 (C-1 major), 101.3 (C-1 minor), 100.7 (PhCH), 81.2 (C-4), 70.4 (C-3 major), 70.3 (C-3 minor), 67.8 [OCH₂CH(O)C(CH₃)₂], 67.5 (C-6), 66.0 (C-5), 61.1 $[OCH_2CH(O)C(CH_3)_2 \text{ major}], 60.6 [OCH_2CH(O) C(CH_3)_2$ minor], 57.3 [OCH₂CH(O)C(CH₃)₂ minor], 57.2 [OCH₂CH(O)C(CH₃)₂ major], 56.2 (C-2 minor), 56.1 (C-2 major), 24.4, 18.6 $[OCH_2CH(O)C(CH_3)_2]$, 23.1 (CH₃CON). HRMS (EI): [M]+[•], found 393.178388. C₂₀H₂₇NO₇ requires 393.178753. Anal. calcd for C₂₀H₂₇NO₇: C, 61.06; H, 6.92; N, 3.56. Found: C, 61.00; H, 6.86; N, 3.61%.

3.8.2.5. 3,4-Epoxybutyl 2-acetamido-(R)-4,6-O-benzylidene-2-deoxy-β-D-glucopyranoside, **28**. Two stereoisomers were obtained. Reaction at 10°C: mp 251–253°C; $[\alpha]_D$ –64.5 (*c* 0.9, DMF); MS (CI): *m*/*z* 380 (100%) [M+H]⁺. ¹H NMR (200 MHz, CDCl₃): δ 7.5– 7.3 (m, 5H, Ph), 6.70 (2d, 1H, J_{2.NH} 4.7 Hz, NH), 5.55 (s, 1H, PhCH), 4.62 (d, J_{1,2} 8.3 Hz, H-1 minor), 4.53 (d, J_{1,2} 8.3 Hz, H-1 major), 4.30 (m, 1H, H-6_e), 3.06 [m, 1H, OCH₂CH₂CH(O)CH₂], 2.81, 2.56 [2m, 2H, OCH₂CH₂CH(O)CH₂], 2.04 (s, 3H, CH₃CON), 1.60 [m, 2H, OCH₂CH₂CH(O)CH₂]. 13 C NMR (50 MHz, DMSO-d₆): δ 169.1 (C=O), 137.8, 128.9, 128.0, 126.4 (Ph), 101.7 (C-1 major), 101.4 (C-1 minor), 100.7 (PhCH), 81.3 (C-4), 70.4 (C-3), 67.9 [OCH2CH2CH(O)-CH₂], 66.0 (C-5), 65.8 (C-6), 56.0 (C-2), 49.2 $[OCH_2CH_2CH(O)CH_2], 46.3 [OCH_2CH_2CH(O)CH_2]$ major], 46.2 [OCH₂CH₂CH(O)CH₂ minor], 32.3 [OCH₂CH₂CH(O)CH₂ major], 32.1 [OCH₂CH₂CH(O)-CH₂ minor], 23.0 (CH₃CON). HRMS (CI): [M+H]⁺, found 380.170892. C₁₉H₂₆NO₇ requires 380.170928. Anal. calcd for C₁₉H₂₅NO₇: C, 60.15; H, 6.64; N, 3.69. Found: C, 60.09; H, 6.66; N, 3.96%.

3.8.3. Epoxidation of 4,6-*O*-benzylidene-3-*O*-protected derivatives

3.8.3.1. (R)-2,3-Epoxypropyl 2-acetamido-3-O-acetyl-(R)-4,6-O-benzylidene-2-deoxy- β -D-glucopyranoside, 29. From 13: Yield 0.50 g (82%); mp 279–280°C; $[\alpha]_{D}$ –64.0 (c 1.0, CH₂Cl₂); MS (CI): m/z 408 (15%) [M+H]⁺. ¹H NMR (200 MHz, CDCl₃): δ 7.5–7.3 (m, 5H, Ph), 5.96, 5.94 (2d, 1H, J_{2,NH} 9.5 Hz, NH), 5.48 (s, 1H, PhCH), 5.21 (m, 1H, H-3), 4.55 (d, J_{1,2} 8.5 Hz, H-1 minor), 4.43 (d, J_{1,2} 8.4 Hz, H-1 major), 4.30 (dd, 1H, J_{5.6e} 4.7 Hz, $J_{6e,6a}$ 10.3 Hz, H-6_e), 3.06 [m, 1H, OCH₂CH(O)CH₂], 2.76 [m, 1H, OCH₂CH(O)C $\underline{H}_{cis}H_{trans}$], 2.64 [dd, J_{trans}] 2.7 Hz, J_{gem} 5.2 Hz, OCH₂CH(O)CH_{cis}H_{trans} major], 2.51 [dd, J_{trans} 2.6 Hz, J_{gem} 4.8 Hz, OCH₂CH(O)-CH_{cis}H_{trans} minor], 2.06 (s, 3H, CH₃COO), 1.97 (s, CH₃CON major), 1.96 (s, CH₃CON minor). ¹³C NMR (50 MHz, CDCl₃): δ 171.3, 170.3 (C=O), 136.9, 129.1, 128.2, 126.0 (Ph), 102.5 (C-1 major), 102.1 (C-1 minor), 101.3 (PhCH), 78.5 (C-4), 71.9 (C-3), 70.7 68.5 [OCH₂CH(O)CH₂ $[OCH_2CH(O)CH_2 minor],$ major], 67.7 (C-6), 66.3 (C-5), 54.3 (C-2), 50.8 $[OCH_2CH(O)CH_2 \text{ minor}],$ 50.4 $[OCH_2CH(O)CH_2]$ major], $[OCH_2CH(O)CH_2]$ 44.2 minor], 43.8 $[OCH_2CH(O)CH_2 major], 23.3$ (CH₃CON), 20.9 (CH₃COO). HRMS (CI): [M+H]⁺, found 408.165674. C₂₀H₂₆NO₈ requires 408.165842. Anal. calcd for C₂₀H₂₅NO₈: C, 58.96; H, 6.19; N, 3.44. Found: C, 58.92; H, 6.13; N, 3.51%.

From 23: An aliquot of compound 23 (0.75 mmol) was acetylated in the usual way with acetic anhydride/pyri-

 $(O)CH_2$

65.67; H, 6.46; N, 3.12%.

dine (1:1) (4 mL). The reaction mixture was kept overnight at room temperature and then poured into ice-water. The precipitate obtained was isolated by filtration and, recrystallized from ethanol-water, gave 29 as only one stereoisomer. Yield 0.28 g (92%); MS (FAB): *m*/*z* 430 (100%) [M+Na]⁺. ¹H NMR (200 MHz, CDCl₃): δ 7.5–7.3 (m, 5H, Ph), 5.95 (d, 1H, J_{2.NH} 9.3 Hz, NH), 5.48 (s, 1H, PhC<u>H</u>), 5.20 (t, 1H, $J_{2,3}=J_{3,4}$ 9.9 Hz, H-3), 4.46 (d, 1H, J_{1,2} 8.4 Hz, H-1), 4.31 (dd, 1H, $J_{5,6e}$ 4.7 Hz, $J_{6e,6a}$ 10.3 Hz, H-6_e), 3.50 (dt, 1H, $J_{4,5}$ = $J_{5,6a}$ 9.6 Hz, $J_{5,6e}$ 4.7 Hz, H-5), 3.08 [m, 1H, OCH₂CH(O)CH₂], 2.76 [m, 1H, OCH₂CH(O)-CH_{cis}H_{trans}], 2.64 [dd, 1H, J_{trans} 2.7 Hz, J_{gem} 5.2 Hz, $OCH_2CH(O)CH_{cis}\underline{H}_{trans}$], 2.06 (s, 3H, CH₃ČOO), 1.97 (s, 3H, CH₃CON). ¹³C NMR (50 MHz, CDCl₃): δ 171.3, 170.3 (C=O), 136.9, 129.1, 128.2, 126.1 (Ph), 102.5 (C-1), 101.3 (PhCH), 78.5 (C-4), 71.8 (C-3), 68.5 [OCH2CH(O)CH2], 67.7 (C-6), 66.4 (C-5), 54.4 (C-2), 50.4 [OCH₂CH(O)CH₂], 43.8 [OCH₂CH(O)CH₂], 23.3 (CH₃CON), 20.9 (CH₃COO).

3.8.3.2. (R)-2,3-Epoxypropyl 2-acetamido-(R)-4,6-Obenzylidene-2-deoxy-3-O-methyl-β-D-glucopyranoside, **30**. Yield 0.27 g (47%); mp 211–212°C; $[\alpha]_D$ +102.2 (*c* 0.5, CH₂Cl₂); MS (CI): m/z 380 (100%) [M+H]⁺. ¹H NMR (200 MHz, CDCl₃): δ 7.5–7.3 (m, 5H, Ph), 5.83 (d, 1H, J_{2.NH} 7.7 Hz, NH), 5.52 (s, 1H, PhCH), 5.05 (d, $J_{1,2}$ 8.2 Hz, H-1 minor), 4.95 (d, $J_{1,2}$ 8.2 Hz, H-1 major), 4.31 (dd, 1H, J_{5,6e} 4.3 Hz, J_{6e,6a} 10.3 Hz, H-6_e), $OCH_2CH(O)CH_2],$ 2.77 3.11 [m, 1H, [m, OCH₂CH(O)CH_{cis}H_{trans}], 2.65 [dd, J_{trans} 2.7 Hz, J_{gem} 5.1 Hz, OCH₂CH(O)CH_{cis}H_{trans} major], 2.55 [dd, J_{trans} 2.6 Hz, J_{gem} 5.2 Hz, OCH₂CH(O)CH_{cis}H_{trans} minor], 2.01 (s, 3H, CH₃CON). ¹³C NMR (50 MHz, CDCl₃): δ 170.1 (C=O), 137.2, 129.0, 128.2, 126.0 (Ph), 101.2 (C-1 major), 101.1 (C-1 minor), 100.6 (PhCH), 82.3 (C-4), 78.4 (C-3), 69.2 [OCH₂CH(O)CH₂], 68.7 (C-6), 65.9 (C-5), 57.6 (C-2), 50.7 [OCH₂CH(O)CH₂ minor], 50.3 [OCH₂CH(O)CH₂ major], 44.2 [OCH₂CH(O)CH₂], 29.7 (CH₃O), 23.7 (CH₃CON). HRMS (CI): [M+H]⁺, found 380.169608. C₁₉H₂₆NO₇ requires 380.170928. Anal. calcd for $C_{19}H_{25}NO_7$: C, 60.15; H, 6.64; N, 3.69. Found: C, 59.90; H, 6.62; N, 3.80%.

3.8.3.3. (R)-2,3-Epoxypropyl 2-acetamido-3-O-benzyl-(*R*)-4,6-*O*-benzylidene-2-deoxy- β -D-glucopyranoside, 31. Yield 0.58 g (85%); mp 263–264°C; $[\alpha]_{D}$ –54.0 (c 1.0, CH₂Cl₂); MS (FAB): m/z 478 (97%) [M+Na]⁺. ¹H NMR (200 MHz, CDCl₃): δ 7.5–7.3 (m, 10H, 2Ph), 5.6-5.5 (m, 2H, NH, PhCH), 4.96 (d, J_{1.2} 8.3 Hz, H-1 minor), 4.9-4.8 (m, H-1 major, PhCHAHBO), 4.62 (d, 1H, J_{gem} 11.9 Hz, PhCH_AH_BO), 4.33 (dd, 1H, $J_{5,6e}$ 4.9 Hz, J_{6e,6a} 10.3 Hz, H-6_e), 3.10 [m, 1H, OCH₂CH(O)-CH₂], 2.76 [m, 1H, OCH₂CH(O)CH_{cis}H_{trans}], 2.64 [dd, J_{trans} 2.7 Hz, J_{gem} 5.2 Hz, OCH₂CH(O)CH_{cis}H_{trans} major], 2.52 [dd, J_{trans} 2.6 Hz, J_{gem} 5.2 Hz, OCH₂CH(O)CH_{cis}H_{trans} minor], 1.89 (s, CH₃CON major), 1.87 (s, CH₃CON minor). ¹³C NMR (50 MHz, CDCl₃): δ 170.5 (C=O), 138.3, 137.3, 129.0, 128.4, 128.3, 128.2, 127.8, 126.0 (2Ph), 101.2 (PhCH), 101.0 (C-1 major), 100.7 (C-1 minor), 82.6 (C-4), 76.5 (C-3),

74.4 (PhCH₂O), 70.8 [OCH₂CH(O)CH₂ minor], 68.9 [OCH₂CH(O)CH₂ major], 68.7 (C-6), 66.0 (C-5), 57.1 (C-2), 50.8 [OCH₂CH(O)CH₂ minor], 50.3 [OCH₂CHmajor], 44.2 $[OCH_2CH(O)CH_2],$ 23.6 (CH₃CON). HRMS (CI): [M+H]⁺, found 456.201254. C₂₅H₃₀NO₇ requires 456.202228. Anal. calcd for

C₂₅H₂₉NO₇: C, 65.92; H, 6.42; N, 3.07. Found: C,

3.8.3.4. (R)-2,3-Epoxypropyl 2-acetamido-(R)-4,6-Obenzylidene-2-deoxy-3-O-tert-butyldimethylsilyl-β-Dglucopyranoside, 32. Yield 0.60 g (83%); mp 125-127°C; $[\alpha]_{\rm D}$ -66.0 (c 1.0, CH₂Cl₂); MS (CI): m/z 480 (100%) $[M+H]^+$. ¹H NMR (200 MHz, CDCl₃): δ 7.5–7.3 (m, 5H, Ph), 5.67 (d, 1H, J_{2,NH} 8.1 Hz, NH), 5.46 (s, 1H, PhCH), 4.98 (d, 1H, J_{1,2} 8.5 Hz, H-1 minor), 4.90 (d, 1H, $J_{1,2}$ 8.5 Hz, H-1 major), 3.11 [m, 1H, OCH₂CH(O)CH₂], 2.76 [m, 1H, OCH₂CH(O)- $OCH_2CH(O)CH_2],$ CH₂CH₂(O)CH₂), 2.70 µm, 1m, CCH₂CH₂(O)² CH_{cis}H_{trans}], 2.66 [dd, J_{trans} 2.7 Hz, J_{gem} 5.2 Hz, OCH₂CH(O)CH_{cis}H_{trans} major], 2.52 [dd, J_{trans} 2.7 Hz, J_{gem} 5.2 Hz, OCH₂CH(O)CH_{cis}H_{trans} minor], 2.01 (s, CH₃CON minor), 1.96 (s, CH₃CON major), 0.79 [s, 9H, $OSi(CH_3)_2C(CH_3)_3]$, -0.01, -0.07 [2s, 6H, OSi(CH₃)₂C(CH₃)₃]. ¹³C NMR (50 MHz, CDCl₃): δ 170.3 (C=O), 137.1, 129.0, 128.1, 126.3 (Ph), 101.8 (PhCH), 100.8 (C-1 major), 100.5 (C-1 minor), 82.4 (C-4), 71.2 (C-3), 70.5 [OCH₂CH(O)CH₂ minor], 68.7 [OCH₂CH(O)CH₂ major, C-6], 66.0 (C-5), 59.1 (C-2), 50.8 [OCH₂CH(O)CH₂ minor], 50.4 [OCH₂CH(O)CH₂ 44.2 $[OCH_2CH(O)CH_2]$ major], minor], 44.1 $[OCH_2CH(O)CH_2 major], 25.7 [OSi(CH_3)_2C(CH_3)_3],$ 23.8 (CH₃CON), 18.1 [OSi(CH₃)₂C(CH₃)₃], -4.1, -5.0 $[OSi(CH_3)_2C(CH_3)_3]$. HRMS (CI): $[M+H]^+$, found 480.241486. C₂₄H₃₈NO₇Si requires 480.241756. Anal. calcd for C₂₄H₃₇NO₇Si: C, 60.10; H, 7.78; N, 2.92. Found: C, 60.14; H, 7.80; N, 3.03%.

3.8.3.5. (*R*)-2,3-Epoxy-2-methylpropyl 2-acetamido-(R)-4,6-O-benzylidene-2-deoxy-3-O-tert-butyldimethylsilyl-β-D-glucopyranoside, 33. Yield 0.56 g (76%); mp 137–139°C; $[\alpha]_D$ –42.5 (c 0.8, CH₂Cl₂); MS (CI): m/z494 (50%) $[M+H]^+$. ¹H NMR (200 MHz, CDCl₃): δ 7.5–7.3 (m, 5H, Ph), 5.64 (d, 1H, J_{2,NH} 8.3 Hz, NH), 5.46 (s, 1H, PhCH), 4.92 (d, J_{1,2} 8.4 Hz, H-1 minor), 4.80 (d, J_{1,2} 8.5 Hz, H-1 major), 4.29 (dd, 1H, J_{5,6e} 4.2 Hz, J_{6e,6a} 10.2 Hz, H-6_e), 2.81, 2.57 [2d, 2H, J_{gem} 5.1 Hz, OCH₂C(CH₃)(O)CH₂], 1.98 (s, 3H, CH₃CON), 1.35 $OCH_2C(CH_3)(O)CH_2$ 1.30 minor], [s, s, OCH₂C(CH₃)(O)CH₂ major], 0.79 [s, 9H, OSi(CH₃)₂- $C(CH_3)_3$], -0.14, -0.07 [2s, 6H, $OSi(CH_3)_2C(CH_3)_3$]. ¹³C NMR (50 MHz, CDCl₃): δ 170.1 (C=O), 137.1, 129.0, 128.1, 126.2 (Ph), 101.8 (PhCH), 101.2 (C-1 major), 100.8 minor), 82.3 (C-1 (C-4), 73.2[OCH₂C(CH₃)(O)CH₂ minor], 71.5 (C-3 major), 71.4 (C-3 minor), 70.6 [OCH₂C(CH₃)(O)CH₂ major], 68.7 (C-6), 66.0 (C-5), 58.9 (C-2 minor), 58.7 (C-2 major), 55.8 $[OCH_2C(CH_3)(O)CH_2], 51.6 [OCH_2C(CH_3)(O)-$ CH₂ minor], 50.9 [OCH₂C(CH₃)(O)CH₂ major], 25.7 $[OSi(CH_3)_2C(CH_3)_3],$ 23.7 (CH₃CON), 18.3 $[CH_2C(CH_3)(O)CH_2], 18.1 [OSi(CH_3)_2C(CH_3)_3], -4.1,$ -5.0 [OSi(CH₃)₂C(CH₃)₃]. HRMS (CI): [M+H]⁺, found 494.258167. $C_{25}H_{40}NO_7Si$ requires 494.257406. Anal. calcd for $C_{25}H_{39}NO_7Si$: C, 60.82; H, 7.96; N, 2.84. Found: C, 60.59; H, 7.80; N, 2.87%.

3.8.3.6. (R)-2,3-Epoxypropyl 2-acetamido-(R)-4,6-Obenzylidene-2-deoxy-2-N-3-O-methylidene-β-D-glucopyranoside, 34. Two stereoisomers were obtained. Yield 0.35 g (62%); mp 198–199°C; $[\alpha]_{\rm D}$ +82.8 (c 1.1, CH₂Cl₂); MS (CI): m/z 378 (15%) [M+H]⁺. ¹H NMR (200 MHz, DMSO-d₆, 80°C): δ 7.6–7.3 (m, 5H, Ph), 5.70 (s, 1H, PhCH), 5.27 (d, 1H, J_{gem} 5.0 Hz, OCH_AH_BN), 4.96, 4.95 (2d, 1H, J_{1.2} 8.0 Hz, H-1), 4.72 (d, 1H, J_{gem} 5.0 Hz, OCH_AH_BN), 3.15 [m, 1H, OCH₂CH(O)CH₂], 2.73, 2.60 [2m, 2H, OCH₂CH(O)-CH₂], 2.14 (s, CH₃CON minor), 2.13 (s, CH₃CON major). ¹³C NMR (50 MHz, CDCl₃): δ 179.5 (C=O), 136.4, 129.2, 128.2, 126.1 (Ph), 103.5 (C-1 major), 103.3 (C-1 minor), 101.5 (PhCH), 82.2 (OCH₂N), 81.8 (C-4), 79.0 (C-3), 71.1 [OCH₂CH(O)CH₂ minor], 70.1 [OCH₂CH(O)CH₂ major], 69.1 (C-5), 68.4 (C-6), 61.9 (C-2 major), 61.6 (C-2 minor), 50.2 [OCH₂CH(O)CH₂ minor], 50.0 $[OCH_2CH(O)CH_2]$ major], 44.3[OCH₂CH(O)CH₂ major], 44.2 [OCH₂CH(O)CH₂ minor], 23.2 (CH₃CON). HRMS (CI): [M+H]⁺, found 378.154586. C₁₉H₂₄NO₇ requires 378.155277. Anal. calcd for C₁₉H₂₃NO₇: C, 60.47; H, 6.14; N, 3.71. Found: C, 60.40; H, 6.05; N, 3.80%.

3.9. Azide ring-opening reaction

3.9.1. (R)-3-Azido-2-hydroxypropyl 2-acetamido-3.4.6tri-O-acetyl-2-deoxy-β-D-glucopyranoside 35. To a solution of compound 19 (0.81 g, 2.0 mmol) in acetonitrile (20 mL), lithium perchlorate (0.43 g, 4.0 mmol) and sodium azide (0.52 g, 8.0 mmol) were added. The mixture was stirred at room temperature for 30 days, with stirring. It was then poured into ice-water, and the precipitate obtained was filtered off, dried, and recrystallized from ethanol. Two stereoisomers were obtained in 87:13 ratio (74% d.e.). Yield 0.58 g (65%); mp 104–105°C; $[\alpha]_{D}$ +55.6 (c 0.9, AcOEt); MS (FAB): m/z469 (100%) $[M+Na]^+$. ¹H NMR (200 MHz, CDCl₃): δ 5.88 (d, 1H, J_{2,NH} 8.8 Hz, NH), 4.63 (d, J_{1,2} 8.4 Hz, H-1 major), 4.61 (d, J_{1,2} 8.3 Hz, H-1 minor), 3.43 [d, J 4.6 Hz, $OCH_2CH(OH)CH_2N_3$ minor], 3.32 [d, J 5.2 Hz, OCH₂CH(OH)CH₂N₃ major], 2.07, 2.02, 2.01, 2.00, 1.94 (4s, 12H, 4CH₃CO). ¹³C NMR (50 MHz, CDCl₃): δ 171.1, 170.9, 170.7, 169.3 (4C=O), 101.8 (C-1), 72.3 [OCH₂CH(OH)CH₂N₃], 72.2, (C-4), 71.9 (C-3), 69.7 [OCH₂CH(OH)CH₂N₃ major], 69.5 [OCH₂CH(OH)-CH₂N₃ minor], 68.3 (C-5), 62.0 (C-6), 54.5 (C-2 minor), 54.3 (C-2 major), 53.1 [OCH₂CH(OH)CH₂N₃ major], 52.8 [OCH₂CH(OH)CH₂N₃ minor], 23.3 (CH₃CON), 20.7, 20.6, 20.5 (3CH₃COO). HRMS (CI): [M+H]⁺, found 447.172302. C₁₇H₂₇N₄O₁₀ requires 447.172719. Anal. calcd for $C_{17}H_{26}N_4O_{10}$: C, 45.74; H, 5.87; N, 12.55. Found: C, 45.61; H, 5.70; N, 12.62%.

3.9.2. (*R*)-**3**-Azido-2-hydroxypropyl 2-acetamido-(*R*)-**4,6-O-benzylidene-2-deoxy-\beta-D-glucopyranoside, 36**. To a solution of compound **35** (0.67 g, 1.5 mmol) in methanol (15 mL) was added a solution of sodium

methoxide (1.0 mmol) in methanol (2 mL). After 30 min at room temperature, the mixture was neutralized by addition of Dowex 50 resin (H⁺ form), filtered, and evaporated. The solid obtained was dissolved in benzaldehyde (10 mL), and zinc chloride (0.5 g) was added. The mixture was stirred for 2 days and then poured into hexane/water (1:1) (50 mL) with stirring. The precipitate was filtered off, washed with water and with hexane, and then recrystallized from ethanol. Yield 0.43 g (70%); mp 228–229; MS (CI): m/z 409 (5%) [M+H]⁺. ¹H NMR (200 MHz, DMSO- d_6): δ 7.86 (d, $J_{2.NH}$ 8.4 Hz, NH minor), 7.83 (d, J_{2,NH} 8.4 Hz, NH major), 7.5-7.3 (m, 5H, Ph), 5.59 (s, 1H, PhCH), 5.28 (m, 2H, 2OH), 4.47 (d, J_{1.2} 7.8 Hz, H-1 minor), 4.44 (d, J_{1.2} 7.8 Hz, H-1 major), 4.19 (dd, 1H, $J_{5,6e}$ 4.3 Hz, $J_{6e,6a}$ 10.1 Hz, H-6_e), 1.1 (s, 3H, CH₃CON). ¹³C NMR (50 MHz, DMSO-d₆): δ 169.3 (C=O), 137.7, 129.9, 128.1, 126.4 (Ph), 102.2 (C-1 major), 102.0 (C-1 minor), 100.7 (PhCH), 81.2 (C-4), 70.4 [OCH2CH(OH)CH2N3], 70.3 (C-3), 68.6 [OCH₂CH(OH)CH₂N₃], 67.8 (C-6), 66.0 (C-5), 56.1 (C-2), 53.2 [OCH₂CH(OH)CH₂N₃], 23.1 (CH₃CON). HRMS (CI): [M+H]⁺, found 409.171958. $C_{18}H_{25}N_4O_7$ requires 409.172325. Anal. calcd for C₁₈H₂₄N₄O₇: C, 52.94; H, 5.92; N, 13.72. Found: C, 52.88; H, 5.87; N, 13.80%.

3.9.3. (R)-2-Acetoxy-3-azidopropyl 2-acetamido-3-Oacetyl-(R)-4,6-O-benzylidene-2-deoxy- β -D-glucopyranoside, 37. Compound 36 (100 mg, 0.25 mmol) was acetylated in the usual way with acetic anhydride/pyridine (1:1) (5 mL). The reaction mixture was kept overnight at room temperature and then poured into ice-water. The precipitate obtained was isolated by filtration and recrystallized from ethanol, giving compound 37 as a white solid. Only one stereoisomer was obtained. Yield 109 mg (89%); mp 218–219°C; $[\alpha]_{\rm D}$ –64.9 (c 1.0, CH₂Cl₂); MS (CI): *m*/*z* 493 (70%) [M+H]⁺. ¹H NMR (200 MHz, CDCl₃): δ 7.5-7.3 (m, 5H, Ph), 5.90 (d, 1H, J_{2,NH} 9.4 Hz, NH), 5.49 (s, 1H, PhCH), 5.20 (t, 1H, $J_{2,3} = J_{3,4}$ 9.9 H-3), Hz, 5.05 [m, 1H. OCH₂CH(OAc)CH₂N₃], 4.40 (d, 1H, J_{1.2} 8.4 Hz, H-1), 4.30 (dd, 1H, $J_{5,6e}$ 4.8 Hz, $J_{6e,6a}$ 10.4 Hz, H-6_e), 4.11 (m, 1H, H-2), 3.41 [d, 2H, J 4.7 Hz, [d, OCH₂CH(OAc)CH₂N₃], 2.08, 2.07, 1.95 (3s, 9H, 3CH₃CO). ¹³C NMR (50 MHz, CDCl₃): δ 171.5, 170.2, 170.1, (3C=O), 136.8, 129.1, 128.3, 126.0 (Ph), 102.0 (C-1), 101.3 (PhCH), 78.5, (C-4), 71.7 (C-3), 70.5 $[OCH_2CH(OAc)CH_2N_3], 68.5 [OCH_2CH(OAc)CH_2N_3],$ 67.0 (C-6), 66.4 (C-5), 54.1 (C-2), 50.5 [OCH₂CH(OAc)CH₂N₃], 23.3 (CH₃CON), 20.9, 20.8, (2CH₃COO). Anal. calcd for C₂₂H₂₈N₄O₉: C, 53.65; H, 5.73; N, 11.38. Found: C, 53.82; H, 5.53; N, 11.12%.

Acknowledgements

We thank the Ministerio de Ciencia y Tecnología (Spain) and the FEDER program for financial support (BQU2001-2425), and José Martín Reina for his collaboration in the preparation of the starting material.

References

- Iglesias-Guerra, F.; Candela, J. I.; Blanco, E.; Alcudia, F.; Vega-Pérez, J. M. *Chirality* 2002, 14, 199–203.
- Rao, A. S.; Paknikar, S. K.; Kirtane, J. K. *Tetrahedron* 1983, 36, 2323–2367.
- Sharpless, K. B.; Behrens, C. H.; Katsuki, T.; Lee, A. W. M.; Martin, V. S.; Takatani, M.; Viti, S. M.; Walker, F. J.; Woodward, S. S. *Pure Appl. Chem.* **1983**, *55*, 589–604.
- 4. Gorzynski-Smith, J. Synthesis 1984, 629-656.
- Montgomery, J. A.; Johnston, T. P.; Shealy, Y. F. In Burgers Medicinal Chemistry; Wolff, M. E., Ed. Drugs for Neoplastic Diseases; John Wiley and Sons: New York, 1979; pp. 595–670.
- Halmos, T.; Santarromana, M.; Antonakis, K.; Scherman, D. Eur. J. Pharmacol. 1996, 318, 477–484.
- Lin, T. S.; Fischer, P. H.; Prusoff, W. H. J. Med. Chem. 1980, 23, 1235–1237.
- Yamashita, M.; Takahashi, C. *Heterocycles* 1993, 36, 651–654.
- 9. Hall, A. G.; Tilby, M. J. Blood Rev. 1992, 6, 163-173.
- 10. Hovinen, J. Acta Chem. Scand. 1996, 50, 1174-1176.
- Iglesias-Guerra, F.; Romero, I.; Alcudia, F.; Vega-Pérez, J. M. Carbohydr. Res. 1998, 308, 57–62.
- Iglesias-Guerra, F.; Candela, J. I.; Bautista, J.; Alcudia, F.; Vega-Pérez, J. M. Carbohydr. Res. 1999, 316, 71–84.
- Vega-Pérez, J. M.; Candela, J. I.; Blanco, E.; Iglesias-Guerra, F. *Tetrahedron* 1999, 55, 9641–9650.
- Vega-Pérez, J. M.; Candela, J. I.; Iglesias-Guerra, F. J. Org. Chem. 1997, 62, 6608–6611.
- 15. Thomas, E. W. Carbohydr. Res. 1970, 13, 225-228.
- 16. Legler, G.; Bause, E. Carbohydr. Res. 1973, 28, 45-52.
- 17. Clarke, A. J.; Strating, H. Carbohydr. Res. 1989, 188, 245–250.
- Bordier, H. P.; Rodriguez, E. B.; Stick, R. V.; Stone, B. A. J. Biol. Chem. 1989, 264, 4939–4947.
- Rodriguez, E. B.; Scally, G. D.; Stick, R. V. Aust. J. Chem. 1990, 43, 1391–1405.
- 20. Curatolo, W. Biochim. Biophys. Acta 1987, 906, 111-136.

- Colombo, D.; Scala, A.; Taino, I. M.; Toma, L.; Ronchetti, F.; Tokuda, H.; Nishino, H.; Nagatsu, A.; Sakakibara, J. *Bioorg. Med. Chem. Lett.* 1996, *6*, 1187– 1190.
- Shirahashi, H.; Morimoto, T.; Nagatsu, A.; Murakami, N.; Tatta, K.; Sakakibara, J.; Tokuda, H.; Nishino, H. *Chem. Pharm. Bull.* 1996, 44, 1404–1406.
- Colombo, D.; Scala, A.; Taino, I. M.; Toma, L.; Ronchetti, F.; Tokuda, H.; Nishino, H.; Nagatsu, A.; Sakakibara, J. *Cancer Lett.* 1998, 123, 83–86.
- 24. Colombo, D.; Ronchetti, F.; Scala, A.; Toma, L. Tetrahedron: Asymmetry 1998, 9, 2113–2119.
- Compostella, F.; Colombo, D.; Ferraboschi, P.; Scala, A.; Toma, L.; Ronchetti, F. *Eur. J. Org. Chem.* 2002, 1429–1435.
- 26. Bellucci, G.; Catelani, G.; Chiappe, C.; D'Andrea, F.; Grigò, G. *Tetrahedron: Asymmetry* **1997**, *8*, 765–773.
- 27. Suhr, R.; Scheel, O.; Thiem, J. J. Carbohydr. Chem. 1998, 17, 937–968.
- 28. Rodebaugh, R.; Fraser-Reid, B. Tetrahedron 1996, 52, 7663–7678.
- 29. Peter, M. G.; Boldt, P.-C.; Petersen, S. Liebigs Ann. Chem. 1992, 1275–1279.
- Iglesias-Guerra, F.; Candela, J. I.; Espartero, J. L.; Vega-Pérez, J. M. *Tetrahedron Lett.* 1994, 35, 5031–5034.
- Vega-Pérez, J. M.; Vega, M.; Blanco, E.; Iglesias-Guerra, F. *Tetrahedron: Asymmetry* 2001, 12, 135–147.
- Vega-Pérez, J. M.; Vega, M.; Blanco, E.; Iglesias-Guerra, F. *Tetrahedron: Asymmetry* 2001, 12, 3189–3203.
- Vega-Pérez, J. M.; Candela, J. I.; Romero, I.; Blanco, E.; Iglesias-Guerra, F. *Eur. J. Org. Chem.* **2000**, 3949–3956.
- Charon, D.; Chaby, R.; Malinvaud, A.; Mondange, M.; Szabó, L. *Biochemistry* 1985, 24, 2736–2742.
- 35. Kiso, M.; Anderson, L. *Carbohydr. Res.* **1979**, *72*, C12–C14.
- Shaban, M. A. E.; Renhold, V. N.; Jeanloz, R. W. Carbohydr. Res. 1977, 55, 213–233.
- 37. Horton, D. Org. Synth. 1973, 5, 1-5.